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Gut microbiota modulates adoptive cell therapy via CD8α dendritic cells and IL-12

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Adoptive T cell therapy (ACT) is a promising new modality for malignancies. Here, we report that adoptive T cell efficacy in tumor-bearing mice is significantly affected by differences in the native composition of the gut microbiome or treatment with antibiotics, or by heterologous fecal transfer. Depletion of bacteria with vancomycin decreased the rate of tumor growth in mice from The Jackson Laboratory receiving ACT, whereas treatment with neomycin and metronidazole had no effect, indicating the role of specific bacteria in host response. Vancomycin treatment induced an increase in systemic CD8α+ DCs, which sustained systemic adoptively transferred antitumor T cells in an IL-12-dependent manner. In subjects undergoing allogeneic hematopoietic cell transplantation, we found that oral vancomycin also increased IL-12 levels. Collectively, our findings demonstrate an important role played by the gut microbiota in the antitumor effectiveness of ACT and suggest potentially new avenues to improve response to ACT by altering the gut microbiota.

Introduction
Infiltration of primary tumors by Th1 and cytotoxic T cells provides one of the strongest prognostic factors for outcome at all stages of disease (1, 2). The importance of T cells for the antitumor response has been recently demonstrated using adoptive transfer of in vitro expanded tumor infiltrating lymphocytes (TIL) or genetically modified T cells. These approaches have shown enormous promise in clinical trials, including overall response rates of 40% in metastatic melanoma, 90% in acute lymphoblastic leukemia, and 40% in chronic lymphoblastic leukemia (3–9). As an alternative strategy to generate autologous antitumor T cells, expansion of tumor-specific T cells from peripheral blood is an attractive approach. A phase I study that evaluated the feasibility of expanding HER-2/neu (HER2) vaccine–primed peripheral blood T cells in vivo resulted in clinical responses in 43% of patients (10). Moreover, several studies have shown that patients can be immunized against tumor-associated antigens (TAA) (11–17), expanding the chances to implement this method. In mice, T cells purified and expanded from periphery after vaccination with TAA enhanced the effects of therapeutic vaccines in weakly immunogenic tumor models (18) or in a B16 melanoma model after a BM transplant (19).

Despite the promising response rates of these approaches, there is still an incidence of treatment failure. The reasons for this are incompletely understood (20) but likely include endogenous barriers such as peripheral tolerance and immune evasion by the tumor. The limited success also raises the question of other, yet to be discovered, regulators of T cell function that are relevant to tumor control.

Several studies have shown the influence of commensal microbes on T cell phenotype and function (21–24). For instance, colonization of germ-free mice gut with a cocktail of bacteria from Clostridiales clusters IV, XIVa, and XVIII is sufficient to drive Treg differentiation (23). Another example is the influence of bacteria or their byproducts on tumor progression in mammary glands, a process mediated by the host immune system (25, 26). Moreover, the gut microbiota has been demonstrated to affect antitumor immune approaches, including immune check-point inhibition (27–29). The translocation of gut bacteria

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to the i.p. space, due to epithelial layer damage, can induce transitory infection with systemic elevation of IL-12 that affects the efficacy of ACT (30). All of these studies provide strong evidence for close and complex interplay between the gut microbiome, tumor development, and antitumor immunotherapies.

DCs are professional antigen presenting cells (APC) that orchestrate various innate and adaptive immune system effector mechanisms and, thus, play a critical role in the antitumor immune response (31–34). The phenotype and function of DCs have been demonstrated to be influenced by the gut microbiota (35).

Here, we examine the relationship between gut microbiome composition and the response to ACT. Indeed, we demonstrate that ACT efficacy is directly influenced by the gut microbiota in a mouse model of cervical cancer. Furthermore, in mice from The Jackson Laboratory (Jax), we show that ACT efficacy is improved by treatment with oral vancomycin, an antibiotic that mostly targets gram-positive bacteria in the gut. Our study indicates that the effect of vancomycin treatment is dependent on an increase of systemic CD8α+ DCs that, through IL-12 expression, sustains systemic adoptively transferred antitumor T cells.

**Results**

*The gut microbiota influences ACT efficacy.* Previous studies have demonstrated that, despite sharing the same genetic background, animals from different vendors harbor different gut microbiota (36). Therefore, we assessed the impact of the gut microbiome on ACT by comparing tumor growth in C57BL/6 female mice from Jax and Harlan Laboratories, now Envigo (Har). Because effective T cells against the E6/E7 HPV proteins can be generated in patients with HPV-associated cancers (12), we developed a model of T cell therapy based on the tissue culture number 1 (TC1) cervix and lung cancer model (37), in which HPV16 E6/E7 proteins are expressed. To study the effects of the gut microbiota on ACT, we obtained 5 × 10^6 CD3+ T cells polarized under Th1 condition — as ACT efficacy depends of cytotoxic activity and IFN-γ production (38) — that were generated by vaccination (39) of donor mice (Jax), and we transferred them to lympho-depleted TC1 tumor–bearing animals (12) 5 days after tumor implantation and 7 days after lympho depletion.

Adoptive T cell transfer influenced tumor progression in both sets of mice (Figure 1A). In Har mice receiving ACT, tumor growth was almost completely abrogated (*P* < 0.001), while in Jax mice, ACT was less effective (*P* < 0.001). Tumor growth differences between Har and Jax receiving ACT were significant (*P* < 0.001). To investigate differences in gut microbiota that could be associated with the observed differences in ACT efficacy, we carried out 16S rRNA marker gene sequencing of stool samples collected at 7 and 21 days after ACT. The fecal bacterial communities from Har and Jax mice differed substantially when compared using unweighted UniFrac distance (*P* < 0.001, PERMANOVA test, Figure 1B). This difference was attributable primarily to a diverse range of Bacteroidetes taxa in Har mice, namely Bacteroides, Parabacteroides, Prevotella, the Rikenellaceae family, and the candidate family Bacteroidales S24-7 (Figure 1C). The Jax mice were dominated by Bacteroidales S24-7; no other Bacteroidetes taxa were detected (Figure 1C). These vendor-specific differences in fecal microbiota were also observed in a separate set of mice from each vendor sampled upon arrival (Supplemental Figure 1; supplemental material available online with this article; https://doi.org/10.1172/jci.insight.94952DS1). The change in microbiome composition from 7–21 days within the vendor groups was significantly smaller than the difference between vendors (*P* = 0.03, PERMANOVA test).

To further investigate the relevance of the gut microbiota on ACT efficacy, we treated Har and Jax mice with different antibiotics, as early germ-free condition can have durable consequences that are not possible to change by the introduction of gut microbiota (40). We orally administered vancomycin, an antibiotic against gram-positive bacteria that is not absorbed; thus, its activity is limited to the intestine (41). Vancomycin was orally administered for 10 days before initiation of ACT (42). In Har and Jax mice receiving vancomycin, the total number of bacterial taxa was significantly decreased at 7 days following ACT (*P* < 0.001, Figure 2A), due primarily to depletion of gram-positive taxa. In addition, vancomycin treatment eliminated gram-negative bacteria of the Bacteroidetes phylum, such that the phylum was completely absent in Jax mice at 7 days after ACT and only Parabacteroides remained in Har mice (Figure 2B). These taxa were absent in both the control and vancomycin groups of Jax mice. Additional taxonomic differences between treatment groups are outlined in Supplemental Figure 2. At 21 days, the similarity between vancomycin-treated mice and non–vancomycin-treated mice from the same vendor increased relative to the 7-day time point, indicating a partial transition back to the vendor-specific microbiome states upon discontinuation of vancomycin (*P* < 0.001, Figure 2C).

Following treatment with vancomycin, tumor progression was reduced in Jax mice (*P* < 0.001, Figure 2D), matching the impact of ACT in Har mice without vancomycin (*P* = 0.42). Har mice receiving vancomycin and ACT exhibited no change relative to those not receiving vancomycin (*P* = 0.45). Although 25 days was chosen...
as the end point facilitating the comparison of immunologic status to controls, trends in tumor progression were consistent at 35 days (Supplemental Figure 3A). Overall survival was significantly improved, as well, with the addition of vancomycin to the ACT treatment in Jax mice (Supplemental Figure 3B). Thus, introduction of a single antibiotic was sufficient to alter tumor progression. Because vancomycin was unable to cross the epithelial barrier (41) and produced substantial alterations in the gut bacterial populations, we looked to specific changes in the gut microbiota that were associated with tumor progression. We identified taxa in the Bacteroidetes and Firmicutes phyla where the changes in abundance or presence/absence were consistent with the changes in tumor progression (Supplemental Figures 4 and 5, respectively). All taxa identified belonged to the order Clostridiales, or to the Bacteroidales S24-7 family. Taken together, these data provide strong evidence that the composition of the gut microbiota influences the efficacy of ACT in controlling tumor progression.

**Efficacy of ACT is increased by vancomycin but not neomycin/metronidazole antibiotic treatment.** The balance of different cohabitants in the intestine is maintained by complex interactions among the commensals and the host. The imbalances produced by vancomycin treatment affected overall gut microbiome homeostasis and induced dysbiosis among gram-positive and -negative commensals (Figure 2B). To investigate the role of gram-positive and -negative bacteria subgroups in the modulation of the immune system and the ACT outcome, Jax TC1 tumor-bearing mice were treated with either vancomycin or neomycin/metronidazole (Neo/Met), an antibiotic cocktail that targets gram-negative bacteria, prior to the adoptive transfer of T
cells. Antibiotic treatment continued for 10 days, beginning 1 day before the mice were challenged. Mice treated with a combination of ACT and vancomycin (Figure 3A) showed a significant decrease in tumor growth compared with mice that received ACT only ($P < 0.001$) and with the control group ($P < 0.001$), confirming the previous data (Figure 2D). Tumor growth in mice treated with the combination ACT and Neo/Met (Figure 3B) was not statistically different from those that received ACT alone ($P = 0.50$) but significantly smaller in comparison with the control group ($P < 0.001$). We confirmed these results with a second model of ACT, where we adoptively transferred nonpolarized CD3+ T cells generated from vaccination. Similar to the results obtained using Th1 polarized T cells, mice treated with ACT using nonpolarized T cells showed a significant decrease in tumor growth compared with control mice ($P < 0.01$), and an ultimate increase of ACT efficacy was observed when vancomycin was added to ACT ($P < 0.001$, Figure 3C). Likewise, tumor growth in mice treated with the combination of adoptively transferred nonpolarized T cells and Neo/Met did not differ from the nonpolarized ACT-only group ($P = 0.73$, Figure 3D). These results suggest that the bacteria population responsible for opposing the effect of ACT in Jax mice reside within the vancomycin-sensitive, and not the Neo/Met-sensitive, population.

Increased ACT efficacy by antibiotic treatment is phenocopied by fecal microbiota transplant. To establish a causal relationship between the gut microbiota and the efficacy of ACT, we transplanted the fecal microbiota from Jax to Har mice. We first depleted the resident gut microbiome of Har recipient mice, following a previously published antibiotic regimen (43, 44). We confirmed the efficacy of the antibiotic treatment by PCR, with 16S rRNA gene primers targeting either gram-positive or gram-negative bacteria (Figure 4A).
Har recipient mice were then gavaged twice with fecal extract obtained from Jax mice (1–2 pellets/recipient). To confirm engraftment, we assayed for the presence of Segmented Filamentous Bacteria, which are absent in Jax mice and present in Har mice (36). As expected, the bacteria were present in Har mice before antibiotic treatment and absent following engraftment from Jax mice (Figure 4B).

Two weeks after gavage, donor and recipient mice were enrolled in the ACT protocol. To track the microbiota composition following transplant, we carried out 16S rRNA marker gene sequencing of stool samples collected at 0 and 7 days after tumor implantation (Figure 4C). On day 0, 2 weeks after gavage, the microbiota composition of Har mice that received the fecal transplant differed from that of the Jax donors (P = 0.02, PERMANOVA test) but nonetheless had greater similarity to Jax donor mice than to native Har mice (P < 0.001, 2-tailed t test of unweighted UniFrac distances between groups Figure 4D). On day 7, the distances between samples from donor and recipient mice decreased significantly relative to day 0 (P < 0.001, Figure 4D). Thus, the microbiota of recipient mice adopted a similar configuration to that of the donors, and this configuration was maintained at the second time point. When vancomycin was administered to Har mice following the Jax microbiota transfer, the Har recipients displayed synergy between ACT and antibiotic treatment (P = 0.02), replicating the results from native Jax mice (P = 0.39, Figure 4E). Thus, the microbiota-dependent response to ACT was successfully transferred between mice, confirming that extrinsic maneuvers could be applied to control the gut microbiota and improve ACT immunotherapy.

Figure 3. The efficacy of ACT is increased by vancomycin but not neomycin/metronidazole antibiotic treatment. Tumor growth of mice treated with ACT, using CD3+ T cells from immunized mice polarized under Th1 conditions, and (A) vancomycin or (B) neomycin/metronidazole (Neo/Met). Tumor growth of mice treated with ACT, using nonpolarized cells from immunized mice, and (C) vancomycin or (D) Neo/Met. Tumor growth data are representative of 3 independent experiments with 5 mice per group. Means ± SEM are shown. Differences in tumor volume were evaluated with linear mixed effects models. **P < 0.01, ***P < 0.001.
Gut microbiota composition influences tumor infiltration and systemic expansion of reactive T cells. Given the importance of T cell infiltration into the tumor for the effectiveness of ACT, we investigated both total T cell and anti–HPV E7 tetramer–specific T cell infiltration using IHC and flow cytometry analysis in Jax mice. CD3+ T cell infiltration was augmented in ACT and vancomycin mice compared with vancomycin (P = 0.01) or ACT-only groups (P = 0.03, Figure 5A). The specificity of this antitumor infiltration was assessed by staining with the HPV E7–specific tetramer. The number of E7-specific CD3+CD8+ T cells was increased in the tumors of mice treated with the combination of ACT and vancomycin compared with ACT and Neo/Met (P = 0.02) or ACT alone (P = 0.02, Figure 5B). To understand if this was due to preferential in vitro expansion of CD8+ T cells, we performed flow cytometry immune phenotyping after the in vitro polarization/expansion (Supplemental Figure 6). The results show that in vitro polarization/expansion, before adoptive transfer, does not preferentially expand CD8+ T cells. Thus, in spite of expansion of both CD4+ and CD8+ subsets, only CD8+ T cells specifically accumulate within the tumor. Host myeloid-derived suppressor cells (MDSC), macrophages, and host CD8+ and CD4+ T cells were analyzed as well in tumor (Supplemental Figure 7) and spleen (Sup-
We next examined whether antibiotic treatment influenced the systemic expansion of reactive T cells. Using an IFN-γ ELISPOT assay, we found that splenocytes from TC1 tumor-bearing mice treated with ACT and vancomycin exhibited an increased response against the E7 49–57 epitope compared with the individual treatments (P = 0.005 for ACT, P < 0.001 for vancomycin) or the combination with ACT and Neo/Met (P < 0.001, Figure 5C). There was no statistically significant difference between mice treated with ACT and those treated with ACT and Neo/Met (P = 0.11). We next measured cell division by staining adoptively transferred T cells with CFSE. T cells were adoptively transferred into TC1 tumor-bearing mice treated with vancomycin and untreated mice. Eight days after ACT, cells were recovered from spleen. T cells that had been adoptively transferred into TC1 tumor-bearing mice treated with vancomycin diluted CSFE more than T cells from untreated TC1 tumor-bearing mice (Figure 5D), confirming — together with the ELISPOT and the tetramer staining — an increased proliferation of the adoptively transferred T cells. Thus, depletion of vancomycin-sensitive populations from the gut microbiota of Jax mice increased systemic reactive T cells and therapy effectiveness.

**Increased expression of genes associated with cytotoxic and Th1 profiles in TC1 tumors treated with ACT and vancomycin.** The efficacy of immunotherapy against established tumors depends on lymphocyte recruitment and expansion, effector function, and local tumor microenvironment remodeling (2). Effector functions include cytotoxicity, mediated by perforin/granzyme pathways, and Th1 cytokines such as IFN-γ and IL-12 (45–47). In order to investigate whether gene expression signatures were altered with the addition of vancomycin, we...
analyzed the profiles of ACT and vancomycin–treated mice compared with ACT alone in TC1 tumors at day 25 after tumor implantation. *Perforin 1, Granzyme b, Il12a, Ifng, Tbx21, Rorc, Il17a, Gata3, and Il4 gene expression levels were determined by quantitative PCR (qPCR; Figure 6A). Mice treated with ACT and vancomycin had increased expression of Granzyme B (*P = 0.04), Perforin 1 (*P = 0.04), Il12a (*P < 0.001), and Ifng (*P = 0.02) (Figure 6B). Expression of Il17a and Il4 was not detectable in any condition. We also observed an increase in the number of transcripts for Granzyme b, Il12a, and Ifng in tumors of animals receiving vancomycin alone compared with untreated mice (data not shown). These results support a model in which the treatment with vancomycin induces tumor microenvironment remodeling more supportive for T cell infiltration and cytolytic activity.

**Vancomycin treatment increases the number of systemic CD8α+ DC and IL-12p70 in serum.** We next investigated whether the heightened T cell response, putatively influenced by the microbiome, could be mediated by DCs. To study this possibility, we phenotyped DCs isolated from spleens of untreated, vancomycin–, or Neo/Met-treated mice. In mice, there are 3 major subtypes of DCs in spleen: myeloid DC, CD8α+ DC, and plasmacytoid DC. CD8α+ DC (CD11c+CD11b−B220−CD8α+), have the ability to cross-present tumor antigens to CD8+ T cells, help to maintain tolerance in the steady state, and — upon activation — are able to produce Th1 cytokines such as IL-12 and IFN-γ (48, 49). We found increased populations of CD8α+ DC in the spleens of vancomycin-treated mice as compared with Neo/Met-treated (***P < 0.001) or untreated mice (**P < 0.01, Figure 7A and Supplemental Figure 9). Upon activation, CD8α+ DC are major producers of IL-12 and are known to stimulate inflammatory responses (50). Total DCs (CD11c+) isolated from vancomycin-treated mice displayed a 15-fold increase in Il12a transcripts compared with DCs from untreated controls. On the other hand, Neo/Met-treated mice showed a 2-fold decrease in Il12a transcripts (Figure 7B). Il12a transcripts from total DCs isolated from the vancomycin-treated group were associated with higher levels of IL-12p70 protein in serum when compared with Neo/Met (P = 0.02) or untreated mice (P = 0.02, Figure 7C).
To demonstrate a similar effect of vancomycin on IL-12p70 cytokine levels in humans, we turned to a cohort of patients undergoing allogeneic hematopoietic cell transplantation for hematologic malignancies at University of Pennsylvania (Philadelphia, PA, USA). In December 2015, we instituted a uniform policy of oral vancomycin prophylaxis against *Clostridium difficile*. This allowed us to compare IL-12p70 levels in patients receiving oral vancomycin with those in historical controls who did not receive oral vancomycin but underwent identical treatment. We observed an increase of IL-12p70 levels in serum when patients were treated with vancomycin (*P* = 0.03, Figure 7D and Supplemental Table 1). This suggested that oral vancomycin administration in humans increased the IL-12 concentration in serum, as we observed in mice.

The gut microbiota can trigger TLR4 signaling and induce IL-12 production systemically after total body irradiation (TBI) (30, 51). To investigate the role of TLR4 in microbiota-induced alterations of ACT efficacy, we tested the effects of vancomycin treatment in *Tlr4*-KO mice. *Tlr4*-KO mice treated with vancomycin presented increased levels of IL-12p70 protein in serum relative to *Tlr4*-KO untreated mice (Supplemental Figure 10), following the same pattern of WT mice. Thus, induction of TLR4 was ruled out as the mechanism leading to increased IL-12 levels under vancomycin treatment.

To further investigate the effects of antibiotic treatment on the antigen-presentation function of DCs, we tested the immune response of freshly isolated T cells from immunized mice against DCs isolated from spleens of mice treated with vancomycin or Neo/Met using IFN-γ ELISPOT. Purified T cells had a higher response when the peptide E749–57 was presented by DCs derived from vancomycin-treated mice compared with untreated
mice ($P=0.03$, Figure 7E) or mice treated with Neo/Met antibiotic cocktail ($P=0.04$). The heightened response of T cells in vancomycin-treated mice was abolished by the addition of IL-12 blocking antibody ($P=0.03$, Figure 7E). Overall, these data suggest that microbiota dysbiosis has a robust impact on the CD8α+ DC subpopulation, which in turn upregulates IL-12p70, leading to an enhanced T cell response.

**Effects of gut microbiota on adoptive transfer therapy depends on IL-12.** To validate the role of IL-12 on microbiota effects on ACT, we compared TC1 tumor growth in C57BL/6 IL-12–KO mice, which received ACT in combination with vancomycin treatment, with mice receiving no antibiotics. Vancomycin treatment did not improve ACT efficacy when the Il12 gene was knocked out, as measured by tumor progression ($P=0.32$, Figure 8A) or T cell infiltration ($P=0.26$, Figure 8B), but ACT maintained its effects ($P<0.001$). In order to confirm the importance of this cytokine in the synergy, we injected a blocking antibody for IL-12 into TC1-bearing mice that received ACT and vancomycin. The positive effect of vancomycin treatment upon tumor growth and T cell infiltration was lost, relative to mice who had not received the blocking antibody ($P=0.02$, Figure 8C; $P<0.001$, Figure 8D). Together, these results demonstrate that IL-12 contributes to the microbiota-associated antitumor effect of vancomycin treatment in ACT.

**Discussion**

In this study, we describe the impact of the gut microbiota composition on ACT and demonstrate that the microbiota can be altered to improve ACT efficacy. Using the HPV E6/7–expressing cervical cancer
TC1 model, we report that ACT in tumor-bearing mice is significantly affected by differences in the native composition of the gut microbiota, treatment with antibiotics, and heterologous fecal transfer. ACT efficacy was increased when the bacterial community composition was altered with vancomycin, but it was unchanged when a combination of neomycin and metronidazole was applied. Improvement in ACT efficacy was associated with an increase of systemic CD8\(^+\) DCs and IL-12p70 levels, and with more effective expansion of adoptively transferred antitumor T cells in the animals receiving vancomycin.

Several recent studies have demonstrated the impact of the gut microbiome on cancer immunotherapies (52). In 2013, Iida et al. reported that the gut microbiota played a role in activating the innate immune response against cancer (29). Following s.c. injection of tumor cells in mice, they observed that CpG-oligonucleotide immunotherapy in combination with platinum chemotherapy had a reduced efficacy after antibiotic treatment with a cocktail of vancomycin, neomycin, and imipenem. Control mice in their study harbored a range of Bacteroidetes and Clostridiales species, similar to Har mice in our experiments, which responded optimally to ACT. They reported that individual antibiotics increased the variability of TNF response but did not measure tumor growth in their model after exposure to individual antibiotics. In contrast to their study, we found that the response to ACT could be modulated by specific antibiotic treatments.

In a different mouse model, Viaud et al. reported associations between the small intestine microbiota and the efficacy of antitumor therapy with cyclophosphamide (CTX) (53). At the level of the gut microbiome composition, they observed modest differences in small intestinal microbiota and no difference in cecal microbiota composition, while we found that the fecal microbiota changed drastically in our context. In an approach similar to ours, Viaud et al. applied several antibiotic regimens to alter the gut microbial composition of tumor-bearing mice. Application of vancomycin abrogated the response to CTX therapy. In their model, the response to CTX therapy was mediated by different bacteria and host immune cell populations than the response to ACT in our experiments. Importantly, they demonstrated that only a specific phenotype of T cells, pathogenic and not naive Th17 T cells, could restore the CTX efficacy when administrated with vancomycin. In our experiment, we utilized both CD3\(^+\) nonpolarized and T cells polarized under Th1 conditions in vitro and detected no Th17 signature in treated tumors. Thus, these differences could account for the different outcome when applying vancomycin.

The effect of specific Bacteroides species on tumor growth in mice treated by CTLA-4 blockade was reported by Vétizou et al. (27). Elimination of the species by broad-spectrum antibiotics eliminated the response to therapy, while introduction of B. fragilis bacteria or associated molecules restored the response phenotype. In our experiments, we observed a range of Bacteroides species in Har mice but not in Jax mice, correlating with response to ACT. However, we observed that the Bacteroides genus was eliminated after treatment with vancomycin, ruling out species like B. fragilis as a mechanism for the observed vancomycin-induced response to ACT. In this study, a number of related species in the uncultured Bacteroidales S24-7 family were found to be associated with reduced tumor growth in the vancomycin-treated groups, representing an area of potential overlap with the results in our study.

The genus Bifidobacterium was a particular point of interest for Sivan et al., who carried out a study on PD-L1 blocking antibody therapy in a mouse model of melanoma (28). In their experiments, Jax mice responded more favorably to therapy than mice purchased from Taconic. In our study of response to ACT, Jax mice performed less favorably than Har mice. Furthermore, the genus Bifidobacterium was not prominent in the 16S rRNA marker gene sequencing results from either Jax or Har mice in our experiments; thus, it was ruled out as a potential explanation for the differential response to ACT. However, both studies underline the critical role of the gut microbiota on DCs in the modulation of the antitumor immune response. In addition to providing direct evidence for the impact of the gut microbiota on antitumor immune response, several studies have demonstrated the relevance of gut bacteria to immune development and homeostasis and to T cell differentiation and function (22, 25, 26, 54–56).

To date, numerous HPV E7 DNA–based vaccine design modifications and approaches (i.e., codon optimization, antigen targeting modification, or coinjection of adjuvants) have been shown to augment HPV antigen–specific CTL responses (57–59). These studies show that antigen-specific immune responses in humans and mice generated by HPV E7 vaccines increases tumor cure rates and long-term memory. In the TC1 model, the elicitation and tumor infiltration by antigen-specific CD8\(^+\) T cells is essential for the observed antitumor effects of the HPV E7 vaccine. In alignment with the TC1 model, our results show a specific increase of CD8\(^+\) T cells within the tumor. Given that the efficacy of T cell immunotherapy against established tumors depends on Th1 polarization and cytotoxicity, via the perforin/granzyme system (60–
also associated with an increase of host CD8+ cells in tumor, probably due to a more conductive microen-
vironment or numbers of residual endogenous T cells and contributes to this phenotype, as well.

DCs are central in T cell activation and functions, and they are also influenced by the host gut microbiota
(64). We found an increase in the CD8α+ DC population in splenic DCs from vancomycin-treated mice. These
results align with previous studies, which — although in a different context — found that DCs mediated the
effects of the microbiota on tumor development enhancing specific effector T cells (28, 53). We also showed
that purified T cells have a greater response when the HPV peptide is presented by DCs derived from vanco-
mycin-treated mice, and the enhanced response was abolished by the addition of IL-12 blocking antibody.

IL-12 is mainly produced by monocyte populations and DCs in response to bacterial products or intra-
cellular pathogens, or upon interaction with activated T cells. Moreover, it is essential for the differentia-
tion, proliferation, and maintenance of Th1 responses that lead to IFN-γ and IL-2 production (65, 66). In
our study, the increase in Il12a transcripts by DCs isolated from vancomycin-treated mice was associated
with an increase in IL-12p70 protein in serum. We confirmed these results comparing the IL-12p70 levels
in serum of human subjects undergoing the same therapy, allogeneic hematopoietic cell transplantation for
the treatment of hematologic malignancies, who received oral vancomycin for prophylaxis against the com-
mon nosocomial pathogen C. difficile. These data, although in a different setting, are encouraging for future
studies, as elevated levels of IL-12 in serum are associated with increased numbers of tumor-infiltrating
CD8+ T cells and CD3+ T cells in colorectal cancer (67). Additionally, administration of vancomycin was
also associated with an increase of host CD8+ cells in tumor, probably due to a more conductive microen-
vironment in tumors of mice treated with ACT and vancomycin.

Moreover, antibiotic treatment did not improve ACT efficacy when either Il-12 gene expression
was knocked out or when IL-12/IL12 receptor was antagonized by blocking antibody administration.
Although the IL-12p40 subunit is shared between IL-12 and IL-23 cytokines, their production and secretion
in response to cancer is regulated differentially. Additionally, their functions are different and even opposite
of one another, as IL-23 is more relevant in the generation of the Th17 response and IL-17 production (68,
69). In our analysis, we could not detect amplification of IL-17a transcripts in tumors treated with ACT and
vancomycin. Taken together, these results demonstrate that IL-12 contributes to the synergistic modulatory
effects of vancomycin on ACT therapy and tumor T cell infiltration.

A study performed in B16 tumor-bearing mice identified TLR4 as one possible mechanism through
which microbiota could enhance the positive effect of TBI upon tumor progression (30). Within 24 hours
from TBI, gastrointestinal epithelia damage induced by radiation causes the gut microbiota to leak into the
i.p. space, with consequent transient infection, induction of TLR4, and increase of systemic IL-12 (30).
Based on this work, more recently Bowers et al. showed that TBI-induced IL-12 augments Tc17 cell–medi-
tated tumor immunity (51). By utilizing Tlr4-KO animals, our results preclude this signaling as a mechanism
responsible for the increased levels of IL-12p70 in vancomycin-treated mice. However, our data do not
exclude other TLRs, NLRs or other damage-associated molecular patterns (DAMPs), as bacteria have a
number of ligands beyond LPS that can be responsible for the observed effects.

Evidently, the response to antitumor therapies in mice is mediated by a complex range of microbes and
immune system interactions. As our study highlights, outcomes in mice are influenced by the tumor model,
the therapeutic design, and the gut microbial composition. Intervention with targeted antibiotic therapies
has produced consistent results within each context and therefore will remain an important method for
altering the composition of complex microbial communities in the gut until specific sets of bacterial species
can be identified and cultured in standard collections. The lack of available strains for Bacteoidales S24-7, a
prominent family in the previous studies cited here and a candidate for differential response to ACT in our
study, highlights the difficulty in moving forward with cultured organisms alone.

A better understanding of the mechanisms by which the gut microbiome regulates the efficacy of ACT
will help to maximize the effectiveness of immunotherapies in the future (70). Moreover, we propose that
the gut microbiome will need to be modulated in a therapy-specific manner. The present study underscores
the importance of the gut bacteria to ACT immunomodulation via IL-12–mediated effects on systemic
expansion and tumor invasion of the adoptively transferred T cells.
Methods

Mouse strains and cell lines. Six- to 8-week-old C57BL/6 (H-2b), IL-12p40−/−KO (strain B6.129S1-I12btm1Jm/J), and TLR4-KO (strain B6B10ScN-Tlr4<lps-del>/JthJ) female mice were purchased from Jax and Har. Har mice were sourced from the company’s Indianapolis (Indiana, USA) facility and were DOCK2 WT. Mice were randomly assigned to different experimental groups before tumor challenge. No animals were excluded from the analysis. Mice from different vendors were kept separately, and procedures were established to avoid microbiota transfer among them; only 1 cage was inside the hood at a time, gloves were removed between cages, and chlorine dioxide–based sterilant (clydoxS) was used for hood and cages decontamination. Autoclavable chews were utilized instead of nestles, and individual autoclaved food and water bottles (not automated watering system with drinking valves) were employed for the experiments. The TC1 cells (ATCC), a cervix and lung tumor cell line transformed by HPV E6 and E7 viral oncogenes, were propagated in 5% CO₂ at 37°C and cultured in RPMI 1640 medium with 2 mM L-glutamine (Corning) supplemented with 10% FBS (GIBCO, 1456821), adjusted to contain 1.1 g/l sodium pyruvate and 10 mM HEPES (Corning), and supplemented with 2 mM non-essential amino acids (GIBCO) and 100 U/ml−1 penicillin and 100 mg/ml−1 streptomycin (Corning). TC1 cells were tested to discard mycoplasma contamination.

Immunization. Six- to 8-week-old C57BL/6 mice (Jax) were immunized 3 times (once a week) with 100 μg control plasmid (pCon) E6-E7 DNA plasmid (39) (encoding for E6 and E7 genes) i.m. and sacrificed 1 week after the last immunization.

Tumor challenge. Six- to 8-week-old C57BL/6 mice (Jax and Har) were irradiated with a total dose of 4 Gy and, 48 hours later, were injected s.c. with 1 × 10⁵ TC1 cells per mouse. Solid tumor development was monitored by caliper measurements every other day from the day they were palpable. Experiments were terminated when control mice tumors reached 1.75–2 cm³ (according to University of Pennsylvania guidelines).

Antibiotic administration. Mice received drinking water containing vancomycin (0.5 g/l), neomycin 1 g/l plus metronidazole 1 g/l (MilliporeSigma), or water alone ad libitum for 10 days. The Neo/Met treatment water was supplemented with 1.5 g/l of dextrose, maltodextrin, and sucralose mix.

Adoptive T cell therapy. Adoptive T cells transfer was performed 5 days after tumor implantation. Single cell suspension of spleen cells from immunized C57BL/6 Jax mice (n = 5) were pooled and used to magnetically isolate the lymphocyte population (CD3⁺ T cells) (MiltenyBiotec, catalog 130-095-130). Cells (2.5 × 10⁶ cells/ml) were seeded into 24-well plate, previously coated with mouse anti-CD3 antibody (clone 17A2, eBioscience), with Th1 polarizing media, composed by rmIL-12 (3.3 ng/ml; Peprotech, catalog 210-12) and mouse anti–IL-4 (10 μg/ml; clone 11B11, BioXCell). On the second day of culture, T cells were split and rmIL-2 (0.6 ng/ml; Peprotech, catalog 212-12) was added to the polarizing media. After 3 days of polarization in vitro, 5 × 10⁶ T cells were adaptively transferred i.v. For ACT with nonpolarized T cells, 5 × 10⁶ of freshly isolated CD3⁺ T cells were adaptively transferred i.v.

Microbial transfer. Antibiotic-treated animals (1 g/l each neomycin sulfate and metronidazole, and 0.5 g/l of vancomycin hydrochloride [MilliporeSigma] were used for 14 days and provided ad libitum) were gavaged with 200 μl of fecal bacteria obtained by homogenizing the full contents of 2 feces pellets from the corresponding donor mice in PBS. Recipient mice were housed in a single cage. Fecal samples were collected from donor and recipient mice, allowing the fecal donor mice to serve as a control group for their respective vendor. Detection of universal bacteria (forward, 5′-AGGAGGTGATCCAACCGCA-3′; reverse, 5′-AACCTGGAGAAGGTCGGGAY-3′) (71) and Segmented Filamentous Bacteria (forward, 5′-GACGCTGAGGATCCAACCGCA-3′; reverse, 5′-GACGCTGAGGATCCAACCGCA-3′) (72) was performed as previously described. Briefly, DNA was isolated from mouse stools at different time points using the QIAamp DNA stool extraction kit (Qiagen, catalog 51504), according to manufacturer’s instructions, and standard PCR was performed. PCR products were analyzed by agarose gel electrophoresis and ethidium bromide (EB) staining.

In vivo IL-12 blocking treatment. Mice received via i.p. injections 3 doses (once a week) of 1 mg of monoclonal blocking antibody (In VivoMAB anti–mIL-12p75, clone R2-9A5, BioXCell).

ELISpot. Ninety-six–well MAIP plates (MilliporeSigma, catalog MAIPS 4510) were coated overnight with a 2.5 μg/ml solution of rat anti–mouse IFN-γ (IgG1, clone R4-6A2, BD Biosciences). Bulk splenocytes were plated at 0.5 × 10⁶ cells/well in triplicate and incubated for 20 hours at 37°C with 2 μg/ml peptide (E7 peptide [RAHYNIVTF]) synthesized by JPT (https://www.jpt.com). For the DC assay, DCs were isolated from splenocytes using mouse CD11c MicroBeads (Milteny Biotec, catalog 130-097-059).
according to manufacturer’s instructions. DCs (0.1× 10⁶) were plated in triplicate and incubated 20 hours at 37°C with 0.5 × 10⁶ CD3⁺ T cells from immunized mice and 2 μg/ml of E7 peptides. To block IL-12 in vitro, 10 μg/ml monoclonal antibody IL-12p75 was added to each well (InVivoMAb anti-mIL-12p75, clone R2-9A5, BioXCell). After incubation, plates were washed with PBS and 0.05% Tween 20 (Bio-Rad) and incubated 3 hours at room temperature with anti–mouse biotin-conjugated anti–IFN-γ antibody (rat IgG1, clone XMG1.2, BD Biosciences); Streptavidin-alkaline phosphatase conjugate (BD Biosciences, catalog 554065) was added for 30 minutes. Plates were developed by adding nitroblue tetrazolium/5-bromo-4-chloro-3-indolyl phosphate (Thermo Fisher Scientific, catalog 34042), and spots were then counted using an automated ELISpot reader (AutoimmunDiagnostika GmbH).

**Immunohistochemical tumor analysis.** Tumors were embedded in OCT medium and immediately snap frozen in dry ice. Sections (6-μm thick) were stained for mouse CD3 (CD3 [Early T cell Marker], Rabbit Monoclonal Antibody, RM-9107-S0, Thermo Fisher Scientific) with hematoxylin as a counterstain. Images of the slides were taken using a Nikon Eclipse TI microscope. Investigators performed blind assessment of IHC sections.

**Flow cytometry.** Cells were subjected to up to 7-parameters flow cytometry on a FACSCanto flow cytometer using BD FACSDiva software (BD Biosciences); data were analyzed using FlowJo version X (Tree Star Inc.). The following monoclonal antibodies against mouse markers were used: PerCP/Cy 5.5–conjugated anti-CD8 (clone 53-6.7, BD Pharmingen), APC-conjugated anti-CD3ε (clone 145-2C11, BD Pharmingen), FITC-conjugated anti-CD4 (clone RM4-5, BioLegend), and Brilliant Violet-510–conjugated CD45 (clone 104, BioLegend). iTAg Tetramer/PE - H-2 Db HPV 16 E7 (RAHYNIVTF) (MBL) was used to detect specifically infiltrating E7 CD3 T cells adaptively transferred to mice.

**LIVE/DEAD Fixable Aqua Dead Cell Stain (Invitrogen)** was used to gate live cells, and CellTrace CFSE was used to track adoptively transferred cells and proliferation (Invitrogen). DC phenotyping was done using the following monoclonal antibodies against mouse markers: PE-Cy7–conjugated CD11c (clone N418 eBioscience), APC-conjugated CD11b (M1-70, BioLegend), PerCP/Cy 5.5–conjugated anti-CD8 (clone 53-6.7, BD Pharmingen), APC Cy7–conjugated B220 (clone RA36B2, BioLegend), FITC-conjugated GR-1 (clone RB6-8C5, BioLegend), PE-conjugated F4/80 (clone BM8, BioLegend).

**Gating strategy.** Lymphocytes were determined by the forward and side scatter profile. T cells were assessed after gating for live cells (and CD45⁺ cells in tumor) with anti-CD3ε and anti-CD8. Percentage of tetramer-HPV–specific T cells was quantified inside CD3ε/CD8α cell subset. DC subset was determined by anti-CD11c and anti-CD11b inside live cells subset. CD8α⁺ DCs were assessed by anti-CD8⁺ staining inside CD11c⁺Cd11b⁻ B220⁺ compartment.

**Mouse IL-12 ELISA.** Mouse IL-12 was measured from serum using a DuoSet ELISA mouse IL-12p70 kit according to the manufacturer’s instruction (R&D Systems, catalog DY419-05).

**Human IL-12 ELISA.** Human IL-12 levels were measured from plasma using a Human IL-12 p70 Quantikine HS ELISA Kit according to the manufacturer’s instruction (R&D Systems, catalog HS120).

**Vancomycin treatment of human patients.** From December 29, 2015, all patients undergoing allogeneic hematopoietic cell transplantation received oral vancomycin suspension of 125 mg twice daily beginning on the day of admission for transplantation and continuing until discharge. Vancomycin administration was performed as part of a unit-wide policy instituted in response to the high incidence (20%) of C. difficile infection occurring during the inpatient stay. Samples from 17 consecutive patients who received vancomycin prophylaxis and had available day-28 plasma aliquots were used. Samples from 20 consecutive patients who did not receive vancomycin prophylaxis (i.e., underwent allogeneic transplantation prior to the institution of our vancomycin prophylaxis policy) were used. All identifiers from human samples were removed before analysis.

**Mouse IL-12 gene expression on DCs.** Mouse Il12a gene expression on DCs was quantified using RT² Profiler PCR Array according to manufacturer’s instructions (Mouse Cancer Inflammation & Immunity Crostalk, Qiagen, catalog 330231 PAMM-1812Z).

**Molecular signature.** The relative quantification of the expression levels of selected genes was carried out by qPCR relative to untreated TC1 tumor controls at day 25 agter tumor implantation, using an ABI PRISM Viia7 (Applied Biosystems). Total RNA from TC1 tumors was extracted using Trizol reagent (Invitrogen) according to the manufacturer’s instructions. The purity of the RNA samples was determined using EB visualization of intact 18S and 28S RNA bands after agarose gel electrophoresis. RNA (2 μg of total) was used for cDNA synthesis with random primers, using high-capacity cDNA reverse transcription kit (Applied Biosystem, catalog 4368814). cDNA (50 ng) was used in each real-time PCR reaction run according to
manufacturer’s instructions. TaqMan gene expression assays used to quantify expression levels of; Il12a (Mm00434169_m1), Granzyme B (Mm00442834_m1), Tbx21 (Mm00450960_m1), Il4 (Mm00445259_m1), Perforin (Mm00812512_m1), Gata3 (Mm00484683_m1), Infg (Mm01168134_m1), Il17a (Mm00439618_m1), Rorc (Mm01261022_m1), Actin β (Mm00607939_s1), and Gapdh (Mm99999915_g1).

Bacterial 16S rRNA gene sequencing. Fecal samples were collected from mice housed in a single cage per study group. DNA was extracted from mice samples with the DNA Stool kit (Qiagen) according to the manufacturer’s instruction. For 16S sequencing analysis, the PCR amplicon was prepared using V4 (5′-TGTCAGCMCCGCTGCTG3′) and V6 (primer 5′-TCACGRCACGCTGACG-3′) primers with barcodes. The PCR reaction was done using Phusion High-Fidelity DNA Polymerase (New England Biolabs). The PCR reaction was done with an initial step at 98°C for 30 seconds, followed by 25 cycles of 10 seconds at 98°C, 20 seconds at 72°C, and 5 minutes at 72°C. The PCR product was purified from excess of primers and DNA contaminants using AMPure XP (Beckman Coulter) according to the manufacturer’s instruction; all samples were pooled in equimolar concentrations. The metagenomic sequence reads were generated on the Roche 454 GS FLX.

Sequence reads were analyzed using the QIIME pipeline version 1.8 (73). Reads were discarded if they did not match the expected DNA barcode and primer sequences, contained more than 2 ambiguous base calls (N [IUPAC symbol for an ambiguous base call]), or if the length of 16S gene sequence was less than 200 bp. Operational taxonomic units were selected by UCLUST (74), using the default parameters in QIIME. Taxonomic assignments were determined by searching the Greengenes 16S database (75), after which taxa were assigned if 2 of the top 3 hits were in agreement (the default method in QIIME). Representative sequences from each operational taxonomic unit (OTU) were aligned with PyNAST (76), and a phylogenetic tree was inferred using FastTree (77). The resultant tree was used to compute unweighted UniFrac distances between each pair of samples (78).

Statistics. Sample sizes were chosen on the basis of pilot experiments and our experience with similar experiments. For tumor progression experiments, we employed linear mixed effects models to evaluate the effect of treatment on tumor size. A likelihood ratio test was used to compare models with and without the treatment term. For other comparisons apart from microbiome sequencing, we used one-way ANOVA to test for an overall difference between groups. If a difference was found in ANOVA, we followed with 2-tailed Student’s t tests using a pooled estimate of the variance. Where multiple comparisons were carried out, we adjusted P values using the Holm-Bonferroni method, which controls for the familywise error rate. All t test P values presented are 2-tailed, *P < 0.05, **P < 0.01, ***P < 0.001. For 16S sequence data, the PERMANOVA test was used to test a null hypothesis of no difference between group centroids based on unweighted UniFrac distance (79). PERMANOVA P values were calculated using 999 random label permutations. To assess the effect of time (a within-subjects variable), time point labels were permuted only within mice.

Study approval. All animal studies were approved by the IACUC and University Laboratory Animal Resources at the University of Pennsylvania. Mice were treated in accordance with University of Pennsylvania guidelines. Patients were collected under a research protocol that was approved by the University of Pennsylvania IRB.

Author contributions
MUH designed and performed the experiments, analyzed the data, and drafted the manuscript. KB performed microbiota bioinformatics analyses and gave suggestions for the manuscript. SR assisted with the microbiota experiments and gave suggestions for the manuscript. SG assisted with the ACT experiments. SP assisted with the DC experiments. CT performed microbiota bioinformatics analyses. AG and SG provided human patient samples. MAM and JLT gave suggestions for the manuscript and financial support. FDB gave suggestions for the manuscript and helped with critical advice on how to correctly perform the microbiome studies. CHR gave suggestions for the manuscript, with critical advice on T polarization protocols and T cell therapy. AF conceived the project, designed the study, supervised project planning and execution, and wrote the manuscript.

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