Circadian phase resetting by a single short-duration light exposure

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**BACKGROUND.** In humans, a single light exposure of 12 minutes and multiple-millisecond light exposures can shift the phase of the circadian pacemaker. We investigated the response of the human circadian pacemaker to a single 15-second or 2-minute light pulse administered during the biological night.

**METHODS.** Twenty-six healthy individuals participated in a 9-day inpatient protocol that included assessment of dim light melatonin onset time (DLMO time) before and after exposure to a single 15-second \((n = 8)\) or 2-minute \((n = 12)\) pulse of bright light (9,500 lux; 4,100 K fluorescent) or control background dim light (<3 lux; \(n = 6\)). Phase shifts were calculated as the difference in clock time between the two phase estimates.

**RESULTS.** Both 15-second and 2-minute exposures induced phase delay shifts [median \((\pm SD)\)] of \(-34.8 \pm 47.2\) minutes and \(-45.4 \pm 28.4\) minutes, respectively, that were significantly \((P = 0.04)\) greater than the control condition (advance shift: \(+22.3 \pm 51.3\) minutes) but were not significantly different from each other. Comparisons with historic data collected under the same conditions confirmed a nonlinear relationship between exposure duration and the magnitude of phase shift.

**CONCLUSIONS.** Our results underscore […]

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Introduction

Light is the strongest environmental time cue for resetting the circadian clock in mammals, including humans (1, 2). Initial studies examining the phase-resetting effects of light used long-duration (5 hours) and high-intensity (~9,500 lux) light (2) because the human circadian system was thought to be less sensitive to light than that of other mammals. While the circadian pacemaker in mice (3), rats (4), hamsters (5), and humans (6, 7) is known to respond to a sequence of millisecond flashes (administered over 5 seconds in rats, over 5–60 minutes in mice and hamsters, and over 60 minutes in humans), a single millisecond flash does not reset the mammalian pacemaker at the intensities that have been tested (3, 8), suggesting that circadian resetting requires integration of the photic signal over a longer exposure interval.

A nonlinear relationship exists between light exposure (LE) duration and the magnitude of circadian phase resetting responses. A single 12-minute pulse of light can shift the human pacemaker 8 times more...
efficiently per minute of exposure than a 240-minute (i.e., 4-hour) pulse (9). Intermittent LEs (e.g., 15 minutes every 75 minutes) over the same total 6.5-hour duration induce an ~2.5-hour phase delay, which is ~75% of the phase-resetting effect of 6.5 hours of continuous light, despite representing only 23% of the total light duration, suggesting integration of the photic signal over the exposure interval (10, 11).

The goal of the current study was to test the hypothesis that single discrete brief light stimuli, shorter than 12 minutes, will induce greater circadian phase shifts than dim ambient lighting. Moreover, we hypothesized that most of the resetting response will be induced by the early part of the LE, consistent with a nonlinear relationship between light duration and phase resetting response (12). Therefore, in the current study, we investigated the magnitude of phase resetting responses in humans exposed to very brief LEs (either 15 seconds or 2 minutes of bright [~9,500 lux] light) administered during the biological night as compared with a dim light (<3 lux) control condition. These phase-resetting responses were compared with historic data (9) that examined light pulses of 12 minutes, 1 hour, 2.5 hours, and 4 hours collected under the same experimental conditions.

Results

**LE-induced phase shifts.** To examine the effects of different short-duration pulses of bright light on circadian phase resetting, we studied 26 healthy individuals (Figure 1) in a 9-day inpatient protocol (Figure 2A) that included assessment of endogenous circadian phase before and after they were exposed to a single 15-second \( (n = 8) \) or 2-minute \( (n = 12) \) pulse of bright light \( (9,500 \text{ lux}; 4,100 \text{ K fluorescent}) \) or to control background dim light \( (<3 \text{ lux}; n = 6) \). LE-induced phase shifts were calculated as the difference in circadian phase assessed before and after LE using constant routine protocols in a 9-day inpatient study (Figure 2, A and B). Circadian phase was assessed as the time at which melatonin secretion began under dim light (i.e., dim light melatonin onset [DLMO]). The DLMO for each individual for each study day under each of the three conditions are shown in Figure 3, A–C; phase shifts are shown for each individual under each condition in Figure 3, D–F. Both the 15-second and 2-minute exposures induced median phase shifts (~34.81
is likely related to the light adaptation of ipRGCs with sustained photic stimulation (15). These results that mediate LE-induced phase shifts declines as the duration of the photic stimulus increases (14). This light-evoked action potentials per photon from intrinsically photosensitive retinal ganglion cells (ipRGCs) responsiveness of the circadian pacemaker (3, 4, 8, 13). A recent report documents that the number of additional phase shift produced by further extension of the light stimulus due to a reduction in photic drive follows a nonlinear pattern. Our results in humans are consistent with results from animal studies activated levels, defined as the sustained drive (12). The rate of decline from the prompt to the sustained are recycled back to a “ready” state, and it is maintained asymptotically marginally above baseline non-depleted into a “used” state, the light drive to the pacemaker becomes limited by the rate at which elements begining of a light pulse, defined as the “prompt response” (12). In this model, as activator elements are greater activation rate of light-sensitive elements (a measure of setting effectiveness of light) at the very

The higher-than-expected phase resetting capacity of the short-duration pulses (≤ 12 min) suggests that most of the phase resetting in a long-duration pulse likely occurs at the onset of the LE. This supports the Kronauer model (12) of the effects of LE on phase resetting in humans, which predicts that there is a greater activation rate of light-sensitive elements (a measure of setting effectiveness of light) at the very beginning of a light pulse, defined as the “prompt response” (12). In this model, as activator elements are depleted into a “used” state, the light drive to the pacemaker becomes limited by the rate at which elements are recycled back to a “ready” state, and it is maintained asymptotically marginally above baseline non-activated levels, defined as the sustained drive (12). The rate of decline from the prompt to the sustained drive follows a nonlinear pattern. Our results in humans are consistent with results from animal studies that showed light-induced phase resetting to be most efficient at the beginning of the LE, with minimal additional phase shift produced by further extension of the light stimulus due to a reduction in photic responsiveness of the circadian pacemaker (3, 4, 8, 13). A recent report documents that the number of light-evoked action potentials per photon from intrinsically photosensitive retinal ganglion cells (ipRGCs) that mediate LE-induced phase shifts declines as the duration of the photic stimulus increases (14). This is likely related to the light adaptation of ipRGCs with sustained photic stimulation (15). These results
provide a cellular explanation for the change in phase resetting efficiency based on the duration of the light pulse as observed in the current study.

Exposure to multiple short pulses (5, 15, and 45 minutes) delivered over a long-exposure interval (e.g., 6.5 hours) has been shown to induce significant phase shifts in humans (10, 11, 16). More recently, a train of sixty 2-millisecond pulses administered every minute over 60 minutes induced ~45 minutes of phase delay (6, 7). Given that the circadian pacemaker can integrate photic signals (3, 8), these multiple-exposure studies cannot isolate the phase shift induced by each individual pulse. Mechanistic studies of phase resetting in animals show that the pacemaker can integrate and respond to pulses presented 60 minutes apart (17) and that the majority of a phase-resetting response occurs at the beginning of the light pulse (18–20). Our current results suggest that humans may also be disproportionately sensitive to the first few seconds of a light stimulus (12).

This study had a number of limitations. Firstly, the limited sample size in each LE condition may have precluded detecting differences in phase shifts between the 15-second and 2-minute exposures. Secondly, the magnitude of the shifts in the short-duration pulses is close to the limit of detection for our study techniques. In the current analyses, we assessed phase shifts over three cycles, consistent with previous studies (21–23); however, the endogenous drift over multiple circadian cycles between assessments may influence the phase shift results compared with results calculated across only one or two circadian cycles (22, 24, 25). Additionally, the studies were performed against a dim light background, which reduces the masking effects of light on the circadian pacemaker but can cause moderate sensitization of the system to subsequent bright LEs (26, 27). Although the mean illuminances appeared to differ between the duration conditions with up to a ~45% difference between the highest mean (9,040 lux, 1 hour condition) and lowest mean intensities (6,226 lux, 2 minute condition), it is highly unlikely that this affected the phase resetting responses. Even the lowest mean intensity was more than 10 times higher than the ~500 lux intensity at which phase resetting responses saturate in studied under conditions that are similar to the current study (28). Moreover, the 1-hour condition had lower phase resetting efficiency than the 15-second and 2-minute conditions, even though the mean intensity of the 1-hour condition was higher than the other two conditions.
These results have important physiological and clinical implications. Light levels equivalent to ordinary room light (~100 lux) induce half-maximal phase shifts compared with bright light (~1,000–10,000 lux) when the stimuli are 6.5 hours long (28). Results from the current study demonstrate that even short-duration exposures (on the order of seconds) can cause significant phase resetting. Taken together, the results warrant future studies to examine the effects of short-duration LEs at indoor intensities on circadian phase resetting. Such inadvertent phase resetting may be an unrecognized risk factor for insomnia, circadian rhythm sleep disorders, and a variety of adverse health effects associated with circadian rhythm disruption. Alternatively, the effectiveness of these short-light stimuli for resetting circadian rhythms may have potential therapeutic value to improve compliance with treatments for circadian misalignment associated with shift-work, jet-lag, and other circadian rhythm sleep disorders.

Methods

Participants and prestudy conditions
Twenty-six young healthy individuals (15 females; mean age ± SD = 23.6 ± 3.3 years; range 19–30 years) completed the protocol (Figure 1) in the Intensive Physiological Monitoring Unit in the Center for Clinical Investigation (CCI) at Brigham and Women’s Hospital. The study included three groups: 15-second (n = 8; 5 female; mean age [±SD], 24.2 ± 4.3 years) or 2-minute (n = 12; 6 female; mean age [±SD], 23.3 ± 3.0 years) LE conditions, and the dim-light control condition (n = 6; 4F; mean age [±SD] 23.8 ± 2.7 years). Data from six individuals who voluntarily withdrew prior to study completion were not included in the analyses. All were healthy by physical, psychological, and ophthalmologic exams. For at least three weeks prior to admission, participants maintained a self-selected, consistent 8-hour sleep/rest/dark schedule that was confirmed with calls to a time- and date-stamped voicemail at bedtime and wake time and, for at least 7 days prior to entering the unit, objective monitoring with actigraphy and photometry (Actiwatch-L, Minimitter Inc.). Throughout screening, participants were asked to refrain from the use of any prescription or nonprescription medications, supplements, recreational drugs, caffeine, alcohol, or nicotine. Compliance was verified by urine and blood toxicology during screening and urine toxicology upon entry to the unit.

Study protocol
Participants were studied for 9 days in an environment free of time cues and were supervised by staff trained not to reveal information about the time of day. The schedule consisted of a 3-day baseline (8 hours/16 hours sleep/wake cycle based on average sleep times in the 7 days prior to study entry), an initial 50-hour constant routine circadian phase assessment (CR1, described below), and a 16-hour wake episode during which the experimental LE was presented (LE day). This was followed by an 8-hour scheduled sleep opportunity, a 30-hour CR (CR2), and then a final 8-hour sleep opportunity, after which time they were discharged (Figure 2A). During the CR procedures (29), participants were asked to remain awake while supervised in constant dim light in a semirecumbent posture, with daily nutritional and fluid intake divided into isocaloric hourly portions.

Lighting conditions and experimental LE
Ambient light conditions. During the first 2.5 baseline days, maximum ambient light during scheduled wake was ~190 lux (~48 μW/cm²) when measured in the horizontal plane at a height of 187 cm and ~88 lux (~23 μW/cm²) when measured in the vertical plane at a height of 137 cm. Midway through day 3, ambient light was decreased to <3 lux maximum (1.5 lux [<0.4 μW/cm² to ~0.1 μW/cm²]) when measured in the vertical plane at 137 cm and maintained at that level for the remainder of the study, except during the brief experimental LEs (see below) and during scheduled sleep opportunities when all lights were turned off (0 lux). The ambient lights were dimmed midway through day 3 in order to assess DLMO time as a phase estimate under entrained conditions (9, 30).

Experimental LE conditions. The LE procedure included a constant posture episode during which the bright light pulse was administered. The constant posture was used to negate confounding from ambulatory conditions on research measurements (9). The constant posture began 5.7 hours after wake time on the LE day and lasted for 4.5 hours (to accommodate the longest exposure of 4 hours) (9). The 4.5-hour constant posture interval was approximately centered within the 16-hour wake episode, therefore starting 5.7 hours after wake and ending 5.8 hours before bedtime on that study day. The LE was centered on the
16-hour wake episode, which occurred 7.9 hours after wake (2.2 hours after the start of the constant posture) for the 15-second and 2-minute pulses, respectively.

During the constant posture, participants remained seated in a specific location of the room and wore clear UVEX glasses (Uvex Winter Optical) to remove UV radiation. During the constant posture interval, when participants remained in dim background light, participants alternated fixed gaze (lasting at least 5 minutes) with an equal interval of “free gaze.” During fixed gaze, participants focused on a target on the wall directly in front of them and during free gaze, participants were allowed to look elsewhere as long as they did not close their eyes or shield them from the light. Light illuminance was measured at every change of gaze, with the sensor placed next to the participant’s eye and pointed at the target during fixed gaze and in the angle of gaze during free gaze. Starting 10 minutes immediately prior to the light pulse, all participants maintained fixed gaze. During bright light pulses > 2 minutes, participants continued with their fixed and free gaze pattern, whereas during light pulses ≤ 2 minutes, participants stared directly above at the ceiling where the lights were located for the duration of the bright light pulse. In the control condition, the same sequence as in the 2-minute exposure was followed. During the 15-second or 2-minute LE within this session, the target corneal illuminance was >9,500 lux (≥8 × 10^6 μW/cm²); for the control condition, background lighting (<3 lux) was maintained. The mean ± SD illuminance achieved during the 15-second pulse was 8,022 ± 1,387 lux; during the 2-minute pulse, it was 6,226 ± 1,546 lux; and during the control condition, it was 0.5 ± 0.2 lux. A research technician was present during the entire LE session to administer the LE, measure and record light readings, and monitor adherence to study procedures.

Technical details of light generation and measurement. Ambient room lighting was generated using ceiling-mounted 4,100 K fluorescent lamps (F96T12/41U/HO/EW, 95W; F32T8/ADV841/A, 32W; F25T8/TL841, 25W; Philips Lighting) with digital ballasts (Hi-Lume 1% and Eco-10 ballasts, Lutron Electronics Co. Inc.) transmitted through a UV-stable filter (Lexan 9030 with prismatic lens, GE Plastics). Routine illuminance and irradiance measures were conducted using an IL1400 radiometer/powermeter with an SEL-033/Y/W or SEL-033/F/W detector, respectively (International Light Inc.).

Outcome measures of melatonin phase

Blood samples were collected every 5–60 minutes throughout the protocol, and the plasma was frozen for later assay. Melatonin concentration was determined by double-antibody radioimmunoassay with the Kennaway G280 antiserum (31) by a laboratory blinded to experimental conditions (Specialty Assay Research Core Laboratory, Brigham and Women’s Hospital). The plasma melatonin intra-assay coefficient of variation (%CV) was 8.5% at 5.85 pg/ml and 8.0% at 25.46 pg/ml, and the interassay %CV was 10.8% at 5.98 pg/ml and 14.5% at 23.07 pg/ml.

The DLMO was used as the endogenous circadian phase marker and calculated as the time at which levels of melatonin rose above the 25% peak-to-trough amplitude threshold of a 3-harmonic waveform fitted to the first melatonin secretory episode during CR1 (30, 32). Once the threshold for each individual’s DLMO was calculated, it was applied to each daily melatonin onset and described by letter (a–g; see Figure 2B).

Historic data

In addition to those participants described above, (i) four male participants (21.0 ± 1.8 years) previously studied in our laboratory using a near-identical protocol under dim light conditions were included in the control condition for the phase shift analysis (22), and (ii) 39 healthy young adults (22.2 ± 3.6 years; 16 female) previously studied in our laboratory using near-identical procedures in a protocol that examined the phase shifts in response to LEs of 12 minutes, 1 hour, 2.5 hours, and 4 hours were reanalyzed to evaluate a broader duration response of circadian phase resetting (9). The current and historical studies were the same in participant screening criteria and experimental conditions, except for the (i) LE duration and timing and (ii) CR duration. The 4-hour LE was scheduled to begin 0.5 hours after the start of the 4.5-hour constant posture session (6.2 hours after wake time) and end at the same time as the LE session in this protocol. The 2.5-hour and 1-hour LEs were each scheduled so that the midpoint of the LE occurred at the same time of the midpoint for the 4-hour LE (8.2 hours after wake time). The timing of the 12-minute LE occurred slightly earlier, with the midpoint 12 minutes earlier than the other groups. CR1 was a 50-hour wake episode (same as the current study) in the 12-minute condition and a 50.2-hour wake episode in the 1-hour, 2.5-hour, and 4-hour conditions. CR2 was a 30-hour wake episode (same as the current study) in the 12-minute condition and a 29.8-hour wake episode in the 1-hour, 2.5-hour, and 4-hour conditions (9).
The mean (± SD) light intensities for the historic conditions were as follows: 12 minutes was 7,669 ± 1,321 lux, 1 hour was 9,040 ± 498 lux, 2.5 hours was 8,927 ± 666 lux, and 4 hours was 8,396 ± 1,604 lux (9).

Data analysis
Data are reported as the median (±SD) unless otherwise noted. All analyses were performed using SAS 9.3 (SAS Institute Inc.). Data from 22 of the 26 current participants were included in the analysis of melatonin phase shifts. One male participant from the control condition was excluded due to significant daytime melatonin secretion such that a reliable DLMO could not be determined. For two participants (n = 1 from each of the two LE groups; both females), post-hoc analysis revealed that the LE was 4.4 hours after DLMO, which was outside the target range (0.1–4.1 hours) for inducing 85% of the maximal delay shift (22), so they were excluded from analysis. DLMO in the second cycle of CR2 could not be assessed in 1 female participant in the 2-minute exposure group due to missing blood samples resulting from i.v. malfunction. Four participants (n = 3 from the 1-hour group and n = 1 from the 2.5-hour group) from the historical cohort were also excluded from the analysis due to inappropriate timing of the LE as per the target range described above.

Phase shifts were calculated as the difference between the DLMO on the second cycle of CR1 (pre-LE phase) and the DLMO on the second cycle of CR2 (post-LE phase) three cycles later (cycles d and g; Figure 2A and B) (21–23). Median melatonin phase shift data for each LE duration group were fit by a 4-parameter logistic model to characterize the relationship between phase shift and light duration (SAS PROC NLMIXED). The 4-parameter logistic model was previously found to best fit the relationship between phase shift and stimulus duration or intensity (9, 28). The equation for the 4-parameter logistic model was as follows: y = (a – c) / [1 + (x/b)^d] + c, where “a” is the estimated response of the system to a bright light pulse of 0-second duration, “b” is the duration at which 50% of the maximal shift is observed, “c” is the asymptotic maximal responsiveness of the system, and “d” is a measure of the steepness of the rising portion of the curve.

Statistics
All statistical analyses were performed using SAS 9.3 (SAS Institute Inc.). Due to the limited sample sizes in each group, phase shift data were analyzed by the nonparametric Kruskal-Wallis one-way ANOVA to assess the differences in median phase shifts between LE conditions. Post hoc multiple comparisons were made using Wilcoxon rank-sum test. All statistical tests were two-tailed with a significance threshold set to P < 0.05.

Study approval
The study was reviewed and approved by the Partners Human Research Committee (Boston, Massachusetts, USA), and participants provided written informed consent prior to study.

Author contributions
JFD, REK, CAC, SWL, and EBK designed research. SAR, MASH, AMC, NS, and JFD performed data collection. SAR, MASH, AMC, NS, JFD, REK, CAC, SWL, and EBK performed data analysis and interpretation. SAR, MASH, AMC, NS, JFD, REK, CAC, SWL, and EBK prepared manuscript.

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