

Supplementary Material for:

Title: Longitudinal analysis of naturally acquired PfEMP1 CIDR domain variant antibodies identifies associations with malaria protection

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Table S1. Antigens tested in multiplex immunoassay.

Original Antigen Labels	Figure Labels	Group	Binding Phenotype	Genome/ Isolate	Domain Class	Name used in figures of Rambathia et al PMID. 30365003
HB3var03	CIDR α 1.4 (a)	A	EPCR	HB3	CIDR α 1	α 1.4 (a)
IT4var7	CIDR α 1.4 (b)	A	EPCR	IT4	CIDR α 1	α 1.4 (b)
X1965_2	CIDR α 1.5a (a)	A	EPCR	1965	CIDR α 1	α 1.5a (a)
ERS010323	CIDR α 1.5a (b)	A	EPCR	GA013	CIDR α 1	α 1.5a (b)
ERS01002	CIDR α 1.5a (c)	A	EPCR	GA014	CIDR α 1	α 1.5a (c)
X198_5	CIDR α 1.5b (a)	A	unknown	1918	CIDR α 1	α 1.5b (a)
X198313	CIDR α 1.5b (b)	A	unknown	1983	CIDR α 1	α 1.5b (b)
HB3var02	CIDR α 1.6a	A	EPCR	HB3	CIDR α 1	α 1.6a
ERS01057	CIDR α 1.6b (a)	A	EPCR	GA018	CIDR α 1	α 1.6b (a)
ERS01003	CIDR α 1.6b (b)	A	EPCR	GA019	CIDR α 1	α 1.6b (b)
X1965_8	CIDR α 1.7 (a)	A	EPCR	1965	CIDR α 1	α 1.7 (a)
X1918_3	CIDR α 1.7 (b)	A	EPCR	1918	CIDR α 1	α 1.7 (b)
ERS01043	CIDR α 1.7 (c)	A	EPCR	GA024	CIDR α 1	α 1.7 (c)
IT4var08	CIDR γ 3	A	unknown	IT4	CIDR γ	γ
HB3var05	CIDR δ (a)	A	unknown	HB3	CIDR δ	δ (a)
HB3var35	CIDR δ (b)	A	unknown	HB3	CIDR δ	δ (b)
IT4var02	CIDR δ (c)	A	unknown	IT4	CIDR δ	δ (c)
IT4var30	CIDR α 2.10	B	CD36	IT4	CIDR α 2-6	α 2.10
IT4var24	CIDR α 2.2	B	CD36	IT4	CIDR α 2-6	α 2.2
IT4var33	CIDR α 2.4	B	CD36	IT4	CIDR α 2-6	α 2.4
IT4var61	CIDR α 2.7	B	CD36	IT4	CIDR α 2-6	α 2.7
IT4var45	CIDR α 2.9	B	CD36	IT4	CIDR α 2-6	α 2.9
DD2var01	CIDR α 3.1 (a)	B	CD36	DD2	CIDR α 2-6	α 3.1 (a)
HB3var27	CIDR α 3.1 (b)	B	CD36	HB3	CIDR α 2-6	α 3.1 (b)
IT4var21	CIDR α 3.1 (c)	B	CD36	IT4	CIDR α 2-6	α 3.1 (c)
IT4var26	CIDR α 3.3	B	CD36	IT4	CIDR α 2-6	α 3.3
IT4var15	CIDR α 3.5	B	CD36	IT4	CIDR α 2-6	α 3.5
IT4var14	CIDR α 5	B	CD36	IT4	CIDR α 2-6	α 5
IT4var12	CIDR α 6	B	CD36	IT4	CIDR α 2-6	α 6
IT4var20	CIDR α 1.1 (a)	B/A	EPCR	IT4	CIDR α 1	α 1.1 (a)
igh_var19	CIDR α 1.1 (b)	B/A	EPCR	IGH	CIDR α 1	α 1.1 (b)
raj116_var	CIDR α 1.1 (c)	B/A	EPCR	raj116	CIDR α 1	α 1.1 (c)
ERS010178_NODE_17	CIDR α 1.8a	B/A	EPCR	GA026	CIDR α 1	α 1.8a
X2053_3	CIDR α 1.8b (a)	B/A	EPCR	GA027	CIDR α 1	α 1.8b (a)
ERS010532_NODE_326	CIDR α 1.8b (c)	B/A	EPCR	GA029	CIDR α 1	α 1.8b (c)
AMA1	AMA1	non-var	N/A	N/A	AMA1	N/A
BSA	BSA	non-var	N/A	N/A	BSA	N/A
CSP	CSP	non-var	N/A	N/A	CSP	N/A
MSP1	MSP1	non-var	N/A	N/A	MSP1	N/A
tetanus toxoid	tetanus toxoid	non-var	N/A	N/A	tetanus	N/A

Listed antigens were used in a multiplex bead-based immunoassay to determine antigen-specific IgG reactivity of plasma from participants in the Kalifabougou cohort. Bovine serum albumin (BSA) and tetanus toxoid were used as controls for non-specific cross-reactivity and the predictable response to tetanus vaccination, respectively.

Table S2. Differential acquisition of CIDR domain class-specific IgG antibodies with age and/or malaria exposure.

Domain Class	n	PfEMP1 group	Binding Phenotype	Slope for domain class	Coefficient (age:domain class interaction term)	Standard error	t	P value	BH-adjusted P value
CIDR γ	340	A	unknown	0.224	0.0485	0.0134	3.63	0.000283	0.00141
CIDR δ	340	A	unknown	0.209	0.0357	0.00796	4.49	<0.0001	<0.0001
CIDR α 1	340	A	EPCR	0.208	0.0682	0.00435	15.7	<0.0001	<0.0001
CIDR α 2-6	340	B	CD36	0.115	-0.0935	0.00446	-21.0	<0.0001	<0.0001

Refers to Figure 3B. To determine CIDR domain classes for which specific IgG was acquired more rapidly than the other variants, the change in variant-specific IgG reactivity with age was compared between all variants within each CIDR domain class and all other variants. Specifically, for each CIDR domain class, a linear regression model was performed for children <8 years of age, which represents the linear portion of the plot. The dependent variable was log-transformed antigen-specific IgG reactivity; the independent variables were presence of *P. falciparum* parasitemia (determined by PCR), age, and PfEMP-1 variant type dichotomized as the PfEMP-1 domain class of interest or variants in all other domain classes with the latter being the reference level. An interaction between age and CIDR class was included in the model. Tabulated coefficient and statistics are for the age:domain class interaction term. P values were adjusted for multiple testing (5 coefficients per model times 4 domain classes) using the Benjamini-Hochberg (BH) method. Non-PfEMP-1 antigens were not included in the analysis.

Table S3. Pairwise comparison of slopes between CIDR classes.

Domain class 1	Domain class 2	F statistic	P value
CIDR α 1	CIDR γ	1.3852	0.2393
CIDR α 2-6	CIDR γ	88.663	< 2.2e-16
CIDR δ	CIDR γ	0.816	0.3665
CIDR α 1	CIDR δ	0.0333	0.8552
CIDR α 2-6	CIDR δ	164.38	< 2.2e-16
CIDR α 2-6	CIDR α 1	415.19	< 2.2e-16

Refers to Figure 3B. To determine if the slopes for IgG reactivity between each CIDR domain classes were significantly different from each other in a pairwise manner, we compared a simple linear model in which the slopes of any two CIDR domain classes were parallel to a more complex linear model that included an interaction between age and CIDR domain class using analysis of variance tables. For these models, the predictor variable was IgG reactivity (as log₁₀ arbitrary units) and independent variables were age, *P. falciparum* PCR status at baseline, and domain class as two-level factor variable. Analysis was limited to children <8 years of age (n = 340). A P value < 0.05 indicates that the slopes for the two domain classes are significantly different.

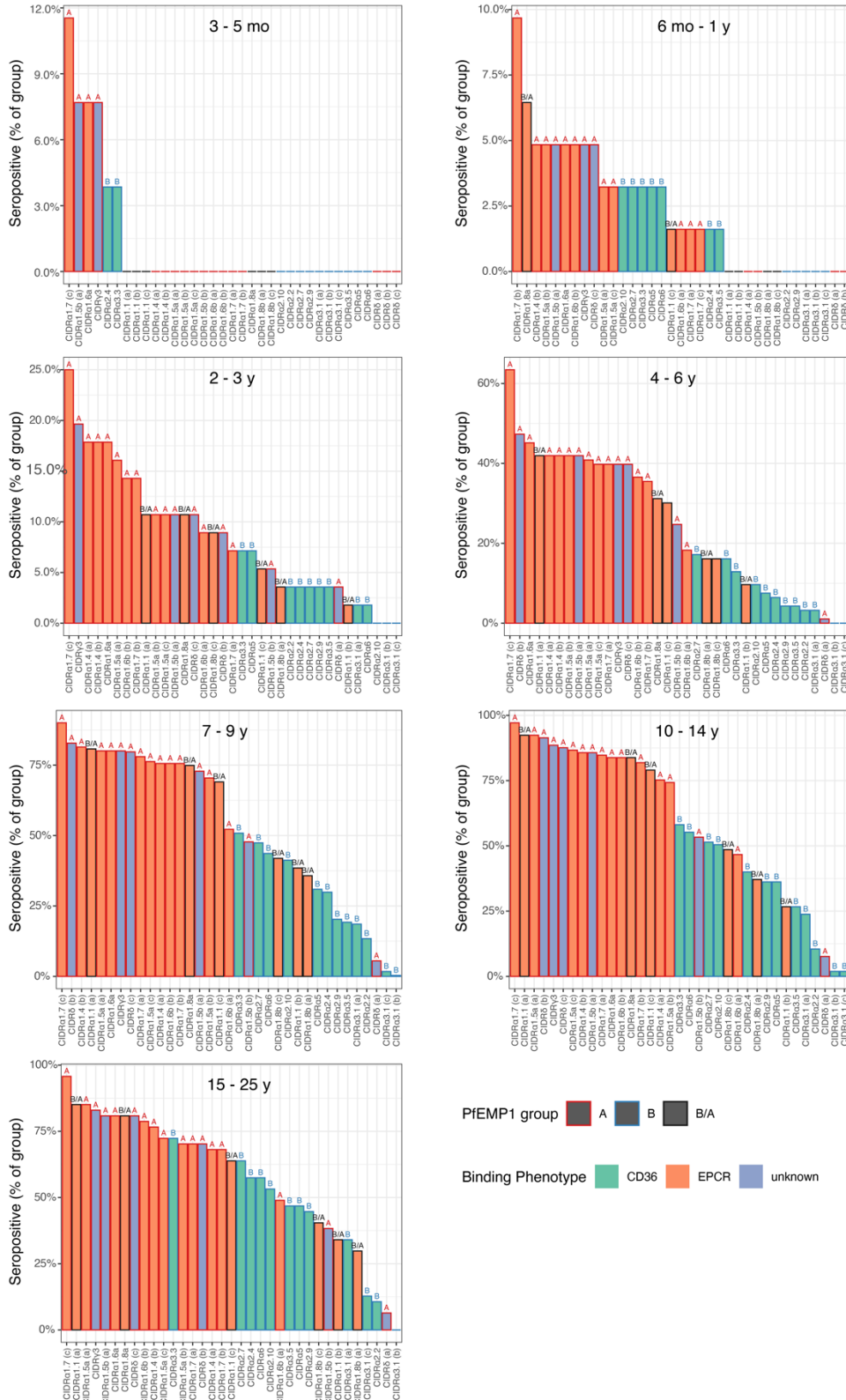


Fig. S1. Antibodies specific for group A and EPCR-binding phenotypes are acquired earlier in life.

Antigens were ranked by seroprevalence to determine the dominant antibody responses for each age group. A response was considered seropositive if the antigen-specific IgG reactivity was greater than the mean reactivity of 20 malaria-naive US donors plus 3 standard deviations.

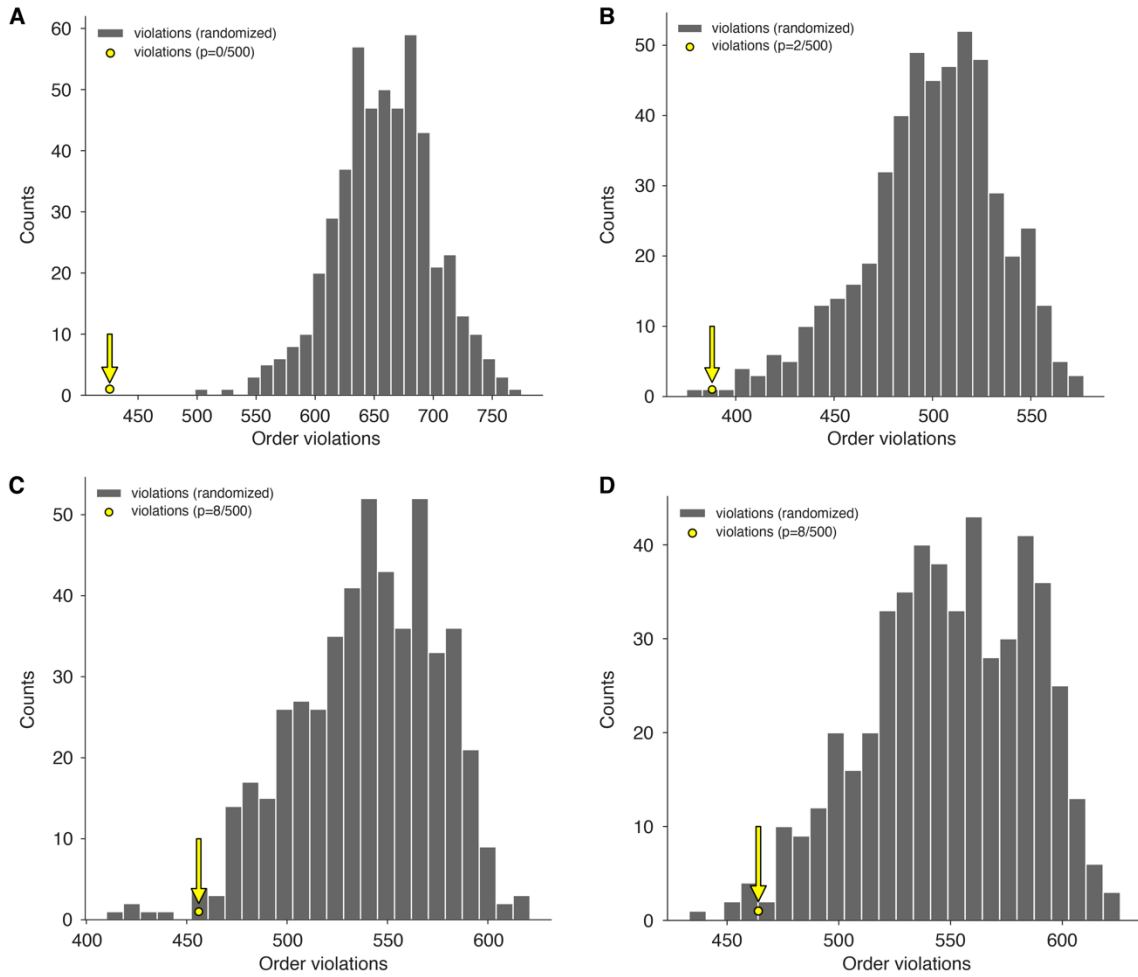


Fig. S2. Significance testing for consensus ordering.

Related to Fig. 5. Observed consensus ordering was compared against 200 independent procedures in which the seroconversion orders for each subject was randomized and consensus ordering was carried out in the same manner, using the total count of consensus-violating seroconversions as a test-statistic (right panel). Fewer consensus violations in real data than in randomized data implies the reported seroconversion ordering is statistically significant. Analysis was performed at the level of **A** individual variants, **B** CIDR domain class, **C** upstream sequence group, and **D** binding phenotype.

Table S4. Relationship between CIDRy3 seropositivity and protection from febrile malaria with inclusion of blood group O as a co-variate.

Covariate	without group O covariate				with group O covariate			
	HR	LCI	UCI	P value	HR	LCI	UCI	P value
Age	1.07	0.998	1.14	0.0582	1.07	0.999	1.14	0.055
CIDRy3	0.411	0.276	0.613	1.26E-05	0.407	0.274	0.605	8.73E-06
group O blood type					0.963	0.678	1.37	0.831
Male	0.858	0.605	1.22	0.39	0.859	0.606	1.22	0.395
presence of HbS allele	0.53	0.311	0.903	0.0196	0.529	0.311	0.902	0.0194

Results of Cox regression models assessing CIDRy3-specific IgG on the risk of febrile malaria after incident *P. falciparum* infection in which covariates were age, gender, presence of the HbS allele, and AMA1-specific IgG reactivity without or with group O blood type. Analysis was restricted to children within the cohort who were at least 6 months of age, began the study negative for *P. falciparum* infection by PCR, and had ABO blood typing performed (218 subjects; 140 malaria events). Malaria risk was determined based on time to clinical malaria, defined as axillary temperature $>37.5^{\circ}$ C and any parasitemia, once parasitemia was detected by PCR. Results are ordered by increasing significance values.