Pressure overload leads to coronary plaque formation, progression, and myocardial events in ApoE−/− mice

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Hypercholesterolemia and hypertension are two major risk factors for coronary artery diseases, which remain the major cause of mortality in the industrialized world. Current animal models of atherosclerosis do not recapitulate coronary plaque disruption, thrombosis, and myocardial infarction occurring in humans. Recently, we demonstrated that exposure of the heart to high pressure, by transverse aortic constriction (TAC), induced coronary lesions in ApoE−/− mice on chow diet. The aim of this study was to characterize the magnitude and location of coronary lesions in ApoE−/− mice after TAC and to assess the susceptibility of coronary plaque to disruption, leading to myocardial events. Here, we describe a reliable pathological condition in mice characterized by the development of coronary lesions and its progression, leading to myocardial infarction; this model better recapitulates human disease. Following TAC surgery, about 90% of ApoE−/− mice developed coronary lesions, especially in the left anterior descending artery, with 59% of the mice manifesting a different magnitude of LAD stenosis. Myocardial events, identified in 74% of the mice, were mainly due to coronary plaque thrombosis and occlusion. That TAC-induced development and progression of coronary lesions in ApoE−/− mice, leading to myocardial events, represents a potentially novel and important tool to investigate the development of coronary lesions and its sequelae in a setting that better resemble human conditions.

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Pressure overload leads to coronary plaque formation, progression, and myocardial events in ApoE−/− mice

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Introduction

Coronary artery disease (CAD) remains the leading cause of morbidity and mortality in the US, accounting for more than 40% of deaths due to cardiovascular disease. Statins have been partially successful, suggesting the need for alternative therapeutic options (1). In this regard, the development of novel therapeutic approaches is impeded, in part, by the use of animal models of atherosclerosis that do not fully mirror the features of human CAD (2).

Since 1994, ApoE−/− mice (3) have been widely used to study the pathogenesis of atherosclerosis and to develop pharmacologic and other interventional therapeutic approaches (4–6). In spite of high plasma cholesterol (6), ApoE−/− mice develop minimal atherosclerosis on a regular diet (3). Thus, to induce atherosclerosis, ApoE−/− mice have been fed with Western-type diet (3), resulting in plasma cholesterol levels ≥1400 mg/dl, a level which is markedly higher than those reported in humans (7). Subsequently, the same approach was used for LDL receptor–knockout (Ldlr−/−) mice (8). ApoE−/− and Ldlr−/− mice fed high-fat diets develop extensive lesions in the aortic root, the aorta, and its principal branches; however, they fail to develop atherosclerosis in the coronary arteries, a major affected site in humans (9). Moreover, in these models coronary plaque progression and its sequelae are not observed.

To overcome this drawback and induce coronary lesions, genetic mouse models were generated by excising the srb1 gene (scavenger receptor class B type I, also known as SR-BI) (10) and the Ldlr gene (11) on an ApoE−/− background. However, long-term high-fat diet was still required to induce coronary atherosclerosis in the Ldlr−/−/ApoE−/− mice and remained not sufficient to elicit myocardial infarction (MI). Indeed, additional hypoxic or mental stress was necessary to cause MI, which occurred in only 40% of
the mice (11). SRB1–/–-ApoE–/– mice developed coronary lesions on regular diet, but they had an incredible short lifetime (8 weeks) due to a complex pathological condition, making this model not amenable to preclinical studies (10). Other attempts have been made to induce coronary lesions in mice, such as the heterozygous mutation of fibrillin-1 gene on an ApoE−/− background (12). Although coronary lesions formed, stroke was the primary cause of death in more than 60% of the mice (12). The loss of endothelial nitric oxide synthase (eNOS) was found to enhance atherosclerosis in Western-type diet–fed ApoE−/− mice and induce a complex cardiovascular disease, with the formation of distal coronary atherosclerosis, spontaneous aortic aneurysms, and dissections (13). The systemic loss of the serine-threonine protein kinase AKT1 also enhanced atherosclerosis in Western diet–fed ApoE−/− mice and was associated to the presence of coronary lesions (14). Thus, clinically relevant events of coronary plaque rupture and thrombosis cannot be systematically studied in existing mouse models, presenting a complex cardiovascular disease and the lack of reliable coronary lesion formation and progression.

Established risk factors for CAD are hypercholesterolemia and high blood pressure (15, 16). In line with clinical findings, transverse aortic constriction (TAC) in ApoE−/− mice, on regular chow diet, induced the development of atherosclerotic lesions in the vascular segments exposed to high pressure, including right carotid artery and coronary arteries, but not in the vessels at low pressure (i.e., left carotid artery and descending aorta) (17). The aim of this study was to define the magnitude and location of coronary lesions in ApoE−/− mice after TAC and to investigate whether the susceptibility of coronary plaque to disruption was reproducible and accompanied by “human-like” events, such as MI and/or sudden death.

Here, we report that TAC-induced coronary atherosclerosis can progress to plaque thrombosis, occlusion, and MI. At 8 weeks after TAC, more than 90% of ApoE−/− mice developed atherosclerotic lesions in the coronary arteries, particularly in the left anterior descending artery (LAD). About 60% of the mice presented LAD stenosis.

When subjected to physical stress conditions (i.e., treadmill), more than 70% of the TAC-operated ApoE−/− mice died and the majority showed striking evidence of previous MI. Systematic and robust histological analysis revealed that MI was caused mainly by coronary occlusion or plaque rupture and/or erosion.

Our study demonstrates, for the first time to our knowledge, that ApoE−/− mice are susceptible to coronary lesion formation and its sequela if chronically exposed to elevated cholesterol (~350 mg/ml) and increased pressure afterload. The formation of coronary lesions in ApoE−/− mice following pressure overload makes them of great experimental value. The combined phenotype (high cholesterol/pressure overload with coronary lesions) provides a baseline against which either detrimental or protective genetic and environmental factors, as well as drugs targeting atherosclerosis, can be investigated.

Results

Characterization and distribution of coronary lesions in ApoE−/− mice at 8 weeks after TAC. TAC surgery was performed on ApoE−/− mice (about 25 g of body weight), leading to transthoracic pressure gradient of circa 60 mmHg (Supplemental Figure 1; supplemental material available online with this article; https://doi.org/10.1172/jci.insight.128220DS1). Different parameters were evaluated at different time points, including magnitude and location of coronary lesions, ECGs, echocardiography, and histology of the hearts (Figure 1A). At 8 weeks after TAC, histological analysis of heart sections from the aortic valve (AV) to the apex was performed to identify the presence of coronary lesions, the degree of stenosis, and the lipid content of the plaques (Figure 1, B–E). The majority of early LAD lesions and its branches (~80%) presented lipid deposition in both intima and media (Figure 1B and Supplemental Figure 1A). Macrophages with different degrees of lipid accumulation were observed in the intima and in the media (Figure 1B, early lesions, arrows and arrowheads, respectively). In 20% of the early lesions, the accumulation of the lipids was limited to the intima or to the media (Supplemental Figure 2A). Just as endothelial cells (ECs), activated smooth muscle cells (SMCs) can express adhesion molecules, such as intercellular adhesion molecule (ICAM) and vascular cell adhesion molecule (VCAM), and therefore can interact and retain monocytes/macrophages into the atherosclerotic artery (18). Interestingly, the initial accumulation of lipids into the coronary wall seems to be different throughout the coronary tree. Specifically, the media of smaller coronary arteries in the apex of the heart (i.e., distal LAD and marginal and diagonal arteries) presented greater lipid accrual compared with the proximal LAD (Supplemental Figure 2B), suggesting a different role of SMCs in distinct coronary artery segments during the initial phase of atherosclerosis. Altogether, these findings suggest similar aspects in the onset of coronary atherosclerosis in mice and humans, including the initial local
accumulation of lipids in the intima and media, neointimal thickening, and recruitment of macrophages, which later develop into foam cells.

The majority of advanced coronary lesions presented a marked neointimal thickening with lipid accumulation and different degree of macrophage infiltration. Figure 1B shows 2 examples of advanced lesions: one causing <50% of luminal stenosis, with pronounced neointimal thickening and infiltrated macrophages (advanced lesion 1), and another presenting >50% of stenosis (advanced lesion 2), with robust macrophage infiltration forming foam cells in the intima (asterisks) as well as few in the media (arrowhead). At the same time point, the overall blood cell counts of sham-operated ApoE−/− and TAC-operated ApoE−/− WT mice were not different (Supplemental Figure 3).

Next, 8 weeks after TAC, we quantified the extent of LAD stenosis in sequential myocardial sections from the AV to the apex (Figure 1, C and D). The highest stenosis was observed along the proximal
coronary vasculature, specifically within 5 mm from the AV (Figure 1D). The lipid plaque composition followed a similar longitudinal profile (Figure 1E).

Based on the analysis of the LAD stenosis, we defined the proximal area of the LAD as a “hot spot” (Figure 1F). It is noteworthy to mention that the hot spot in this murine model of coronary lesions is similar to the one observed in human coronary arteries (19).

As expected, cholesterol levels in ApoE–/– mice were significantly higher than those in WT mice (6) and were not different between sham- and TAC-operated ApoE –/– mice (Figure 1G), suggesting that the high pressure following aortic banding, in combination with moderate high cholesterol (~350 mg/dl), is responsible for coronary plaque formation. The analysis of coronary lesions in the hearts of 34 TAC-operated ApoE–/– mice revealed that 59% developed LAD stenosis, 35% showed early lesions without stenosis, and only 6% had no evidence of atherosclerosis (Figure 1H).
Evidence of MI in ApoE–/– mice exposed to physical stress after TAC. By 10 weeks after TAC, more than 60% of ApoE–/– mice had died. Postmortem examination of some of these mice revealed areas of pale myocardium, suggestive of MI.

Next, ApoE–/– mice were physically stressed by treadmill running (once per week), starting at 8 weeks after TAC, when coronary lesions were established. During this protocol, echocardiographic analysis and ECG telemetry were performed (Figure 2A). After 25 weeks, 17% of the TAC-operated ApoE–/– mice survived, 17% died before the physical stress protocol, and 66% died during the physical stress protocol (Figure 2B). The Kaplan-Meier curve in Figure 2C shows that 50% of the ApoE–/– mice died by week 14 and about 80% of the mice died by week 25. Postmortem analysis of these mice also revealed pale myocardial areas suggestive of infarct (Figure 2D).

Eight TAC-operated ApoE–/– mice were implanted with radiotelemetry devices to measure single-lead ECG once per week, at rest, during, and 48 hours after physical exercise. ECG traces showed a clear ST elevation in 3 mice, and arrhythmic events in the remaining. The latter included accelerated idioventricular rhythm, which is associated with myocardial ischemia, and sudden profound bradycardia (sinus bradycardia and atrioventricular block). In the ECG depicted in Figure 2E, the ApoE–/– mouse at 9 weeks after TAC showed ST elevation at rest, which became more prominent during exercise, and the mouse died immediately after running with associated sudden bradycardia.

Masson's trichrome staining of banded hearts from mice that died with ST elevations revealed an extended cardiac fibrosis due to infarction damage. As controls, the Masson's trichrome staining was performed on the banded hearts of WT (Figure 2F, left) and surviving ApoE–/– (Figure 2F, middle) mice at the same time point after TAC, showing some degree of fibrosis, as expected in pressure-overloaded hearts, but substantially less than the infarcted ApoE–/– hearts (Figure 2F, right).

Finally, as expected fractional shortening was decreased following TAC (8–25 weeks, Figure 2G). Echocardiographic images showed left ventricular dilation, reduced function, and evident ventricular wall thinning in infarcted hearts (Figure 2H) compared with noninfarcted hearts (Figure 2I) of TAC-operated ApoE–/– mice.

Evidence of plaque disruption, thrombosis, and MI TAC-operated ApoE–/– mice. Hearts of TAC-operated ApoE–/– mice that died during the physical stress protocol were systematically sectioned from the AV to the apex and stained with H&E. This first histological examination allowed us to identify points of plaque occlusion or disruption for further analysis. H&E staining of sequential myocardial sections revealed 2 severe atherosclerotic lesions in the LAD at 2.95 mm from the AV (Figure 3A, image 4, asterisks, and Figure 3, B–G). Juxtaposed to the right plaque, in Figure 3A, inset i, there are noticeable “white” thrombi (arrowheads) and red blood cells confined within the narrow lumen. Next, we sought to determine whether plaque rupture and/or erosion caused the thrombotic event. Thus, myocardial sections, upstream and downstream to the one shown in image 4 (Figure 3A), were stained for CD41 to identify platelets aggregates, α-smooth muscle actin (αSMA) for SMC, and collagen I (Col I, marker of extracellular matrix) to define the fibrous cap of the plaque (Figure 3C). The immunofluorescent staining of the LAD showed a thin but continuous fibrous cap of the left plaque (Figure 3C, i) and 2 points of rupture in the fibrous cap of the right plaque, evidenced by the interrupted Col I staining (Figure 3C, ii). In Figure 3D, CD41 staining (red) of a consecutive section, confirmed the presence of a thrombus, which extended from the lumen to inside the ruptured plaque (Figure 3D, ii). These events are also depicted in the image in Figure 3E. It appears also clear that the thrombus is confined to the lumen in front of the left plaque (Figure 3, D and F, and relative i images) with thin and undisrupted fibrous cap. Perhaps erosion might have occurred as the thrombus juxtaposes to the luminal surface of the left plaque.

In distal myocardial sections, the thrombus further extended into the ruptured plaque in intimate contact with the thrombogenic material (Figure 3, H and I as well as G and J). Moreover, CD31 staining (ECs) showed the lack of the endothelium on the right-ruptured plaque, while a positive staining can be observed in the upper-left part of the lumen (Figure 3I). Masson's trichrome staining of cross sections from the medium and apex of the heart revealed some perivascular and interstitial fibrosis but no signs of chronic MI (Figure 3K).

Another example of coronary plaque rupture is shown in Supplemental Figure 4. It is noteworthy to mention that platelet aggregation is not present in proximal or distal myocardial sections to 2.95 mm from AV, suggesting that the observed thrombus did not result from stagnant blood flow in the coronary vasculature. Masson's trichrome demonstrated minimal cardiac fibrosis, suggesting the onset of an acute MI due to plaque rupture (Supplemental Figure 4P).
A significant cause of acute coronary syndrome is superficial plaque erosion (20). This event involves fibrous, rather than lipid-rich plaque, and disturbed flow may contribute to endothelial prothrombotic phenotype and apoptosis (21, 22).

H&E staining of sequential myocardial sections of a TAC-operated ApoE–/– mouse that died during physical stress protocol showed the presence of a “white” thrombus at about 3 mm from the AV (Figure 4, A and B). Immunofluorescent staining of consecutive myocardial sections for CD41 corroborated this observation. Whereas at 2.57 mm from the AV — upstream the point of erosion — no thrombus was observed (Figure 4, C–E), at about 3 mm from AV, CD41 staining evidenced the presence of thrombus in the lumen, adjacent to, but not inside the plaque (Figure 4, F and G). This observation was supported by Col I staining showing a thick fibrous cap, thus excluding the rupture of the plaque (Figure 4, H, I, and K). The lack of notable EC nuclei in Figure 4, H and I, suggested...
erosion of the endothelium. CD31 staining confirmed the absence of the endothelium on the plaque in contact with the thrombus (Figure 4J, ii, arrowheads), whereas positive CD31 staining was present on the opposite site of the LAD (Figure 4J, i, arrows, and K), where the thrombus was absent. These data suggest that coronary plaque erosion triggered the formation of thrombus; contributing to MI. Masson’s trichrome staining of the heart with minimal fibrosis suggests the onset of an acute MI due to erosion rather than chronic infarct (Figure 4L).

Evidence of MI caused by coronary occlusion in TAC-operated ApoE−/− mice. Histological analysis of the heart showed coronary stenosis atherosclerosis and no evidence of plaque disruption and thrombosis (Figure 5A).

Specifically, the occlusion of LAD and/or its branches was evident (i.e., diagonal and marginal arteries). H&E, Masson’s trichrome, and Oil Red O staining of myocardial sections from the base to apex revealed the occlusion of the distal LAD and diagonal arteries (Figure 5, B–D). These advanced plaques were filled with cholesterol crystals (Figure 5C, i and ii) and lipids (Figure 5D). Five of the six −TAC-operated ApoE−/− mice presenting with coronary occlusions were also evaluated with ECG telemetry during the physical stress protocol.
Two mice showed ST elevation, and three mice manifested an accelerated idioventricular rhythm, suggestive of myocardial ischemia.

Figure 5E shows the ECG changes of an ApoE–/– mouse at 11 weeks after TAC at rest and during the run on the treadmill. ST elevation was evident already at 46 seconds of running (Figure 5E). After 2 minutes of running, the QRS complex became wider, finally leading to an accelerated idioventricular rhythm and death. In addition to the coronary occlusion, coronary spasm could also have contributed to the myocardial event.

Interestingly, Masson’s trichrome staining of the hearts of all 6 mice that died during the physical stress protocol and presenting with coronary occlusion showed evidence of extended fibrosis (Figure 5F), suggesting chronic MIs, which might have contributed to stress-induced death from ischemic cardiomyopathy.

Evidence of coronary embolism in TAC-operated ApoE–/– mice. Coronary artery embolism is a rare nonatherosclerotic cause of acute MI, attributed mainly to atrial fibrillation, followed by dilated cardiomyopathy and valvular heart disease (23). In most cases, coronary artery embolism involves the distal LAD, causing transmural MI (24). Interestingly, we found a few cases of abrupt occlusion of coronary arteries without significant atherosclerosis. H&E staining of myocardial sections from the base to the apex clearly showed the absence of atherosclerotic plaques, with abrupt interruption of blood flow in the distal LAD at approximately 6 mm from the AV (Figure 6, A and B). It is possible to notice red blood cells in image 7 and the thrombotic occlusion in the distal image 8 of Figure 6A. CD41 immunofluorescent staining displayed platelet aggregation occluding the entire lumen and disrupting the continuity of the endothelium in some
areas (Figure 6, F–I) but not upstream the thrombus (Figure 6, C–E). In Figure 6J, Masson’s trichrome staining revealed endocardial fibrosis and left ventricular dilation, which may have contributed to the formation of thrombosis that caused the coronary embolism. Because of the absence of significant atherosclerosis in this mouse and the presence of thrombus creating an abrupt occlusion of the distal LAD, we classified this case as coronary embolism.

**Causes of MI in TAC-operated ApoE−/− mice subjected to the physical stress protocol.** In addition to systematic histological analysis, ECG, and echocardiographic measurements, we also performed electron microscopy on the pale myocardial tissue from TAC-operated ApoE−/− mice that died during the physical stress protocol. As control hearts, we used ApoE−/− (with no sign of MI) and C57BL/6 mice at the same time point after TAC. Mitochondria in control hearts had preserved morphology, with intact double membrane, compact and organized cristae, and homogenous dense matrix (Supplemental Figure 5A). On the contrary, infarcted hearts showed the typical mitochondrial ultrastructural hallmarks of irreversible injury, including cell swelling, loss of mitochondrial double membrane, and fragmented reticular mitochondria, with distended and disorganized cristae and dishomogeneous matrix (Supplemental Figure 5, B–D), supporting the occurrence of MI.

Based on all this evidence, we found that 74% of the TAC-operated ApoE−/− mice that died during the physical stress protocol showed evidence of MI (Figure 7A). In 17% of the cases we were not able to collect the hearts because the carcasses were removed from the cages. The remaining 9% of the mice had no evidence of coronary atherosclerosis and MI.
We were able to define and quantify the causes of MI as follows: 47% of the mice presented clear evidence of coronary thrombosis, due to plaque rupture (34%) and plaque erosion (13%), and 40% of the mice showed plaque occlusion, with evidence of chronic myocardial fibrosis (Figure 7B). Two mice had abrupt occlusion of coronary arteries by thrombus with no evidence of atherosclerosis, which was therefore classified as coronary embolism (13%).

Discussion

For long time, establishing a mouse model of coronary atherosclerosis and its progression has been the “holy grail” in atherosclerosis research. ApoE−/− and Ldlr−/− mice have been extensively used to study the pathogenesis of atherosclerosis (25) and provided great insights into the mechanisms of human disease. Studies in mice identified different genes involved in atherogenesis, and some of them have been confirmed in human pathology, including genes involved in triglyceride-rich lipoprotein metabolism (26, 27) and PCSK9 (protein convertase subtilisin/kexin type 9) (28, 29). Mouse models have proven to be very valuable to study disease mechanisms; however, many limitations exist that need to be taken into account, especially considering the challenges in translating the biology of atherosclerosis.

To develop atherosclerotic lesions, mice require cholesterol levels (~2000 mg) that far exceed those commonly present in humans (9). More importantly, murine atherosclerosis develops in the aorta and its proximal branches, whereas coronary arteries are typically spared. The hemodynamics of the aorta and coronary arteries, as well as the origins of their SMCs, differ dramatically. SMCs of coronary arteries originate from the proepicardial organ, while the ones of descending aorta arise from somatic precursors (30, 31). Furthermore, plaque rupture, erosion, and thrombus formation, which are the most critical complications of human atherosclerosis, are not present in current models (25, 32, 33). Whereas patients are treated at late stage of the disease when symptoms arise, mechanisms studied in mice focus on the development of atherosclerosis more than its progression. The difficulties in translating some of the studies in atherosclerosis has been ascribed, at least in part, to the aforementioned limitations of current mouse models (4, 9, 34, 35). To date, few attempts have been made to “model” coronary atherosclerosis and its sequelae in mice (10, 11), without realistic success.

Our study identifies the experimental conditions to induce the formation of coronary atherosclerosis and myocardial events by combining moderated hypercholesterolemia (ApoE−/− mice on chow diet) with...
increased afterload (TAC surgery), two major risk factors for CAD (36–39) and its complications. We have systematically characterized the topography, the magnitude, and lipid composition of coronary plaques, in particular of the LAD (Figure 1). The major finding of our study is the formation of coronary lesions in mice exposed to specific pathological conditions (high cholesterol and pressure), which can progress over time and lead to MI under physical stress conditions (treadmill run), mainly due to coronary disruption and thrombosis and coronary occlusion (Figure 7). Physical exertion (40) and emotional stress may act as external triggers for acute MI (41). Similarly, in TAC-operated ApoE–/– mice, physical stress induced myocardial events, in most of the mice (74%), during or within 48 hours from the treadmill run.

Most emphasis has been placed on the role of endothelium and macrophages in the initiation and progression of early atherosclerosis. However, human autopsy studies (42, 43) and in vivo data suggest an important role of SMCs in the initial phase of atherosclerosis. As macrophages, SMCs are known to express a variety of cholesterol uptake receptors, including LDL receptor (44), CD36 (45), and VLDL receptor (46), and, therefore, can form foam-like cells and participate in atherogenesis. SMCs are also able to express adhesion molecules and retain inflammatory cells in the vascular wall. In early coronary lesions of ApoE–/– mice at 8 weeks after TAC, in addition to macrophages and ECs, lipid accumulation was found in SMCs of the media (Figure 1), suggesting an active role of coronary SMCs in the development of atherosclerosis in mice. Interestingly, as in humans, early coronary atherosclerosis was tightly associated with neointimal proliferation.

ApoE–/– mice on high-fat diet showed evidence of buried plaques in the innominate artery (47, 48) but no signs of the coronary lesions and disruption (49, 50), which is the cause of about 75% of coronary thrombi formation in humans (51, 52). Our study is the first to our knowledge demonstrating coronary plaque formation and progression toward disruption (rupture and/or erosion) and occlusion, which were accountable for 87% of the MIs.

It is generally accepted that macrophages play an important role in the progression of atherosclerosis, by contributing as an important source of enzymes catabolizing collagen, a key component of the fibrous cap of the plaque (9, 53, 54). Interestingly, immunofluorescent staining (CD68 for macrophages and αSMA for SMCs) evidenced a robust accumulation of macrophages in the ruptured plaque and specifically adjacent to the points of break of the thin fibrous cap (Supplemental Figure 4E). We are aware that these lineage markers for SMC (αSMA) and macrophages (CD68) are promiscuous (55), as some cells are both, CD68 and αSMA positive. The aim of these stainings was not to quantitatively distinguish between these 2 populations but to show the presence of macrophages in the ruptured plaques. Interestingly, in Supplemental Figure 4, it is noticeable that the majority of CD68 and αSMA staining does not colocalize, but only a minor population is double positive. Therefore, even considering these limitations, our data support the abundance of macrophages in the ruptured plaque, in correspondence to the thinning of the fibrous cap and breaking points.

In 40% of the mice studied, we found evidence of coronary ischemia based on ECG monitoring that revealed ST elevation in some cases and arrhythmic events, such as accelerated idioventricular rhythm, in others. Indeed, we found stenosis and occlusions of the LAD arteries as well as other major coronary arteries (diagonal and marginal) in our mouse model. It is possible that, under physical stress, demand-related ischemia due to CAD and prior infarctions resulted in demise. Some mice also demonstrated acute MI potentially triggered by increased shear stress from the exercise. Prolonged ischemia has been shown to induce vasoconstriction in animals and humans (56–58).

Finally, in a small percentage of the infarcted hearts, we found no evidence of coronary atherosclerosis with an abrupt interruption of blood flow by thrombus (Figure 6). In humans, coronary artery embolism occurs at low frequency and is commonly caused by atrial fibrillation (23, 59). Interestingly, ApoE–/– mice following TAC presented a low rate of coronary embolism, and we did not observe atrial fibrillation in our mice. We do not know the origin of the embolus, but it is possible that the left ventricular dysfunction in our model contributed to formation of left ventricular thrombus.

Interestingly, when the same transstenotic gradient was applied to chow diet–fed Ldlr–/– mice, known to have a milder increase in cholesterol levels than ApoE–/– mice (60), coronary lesions developed at a later time point (Supplemental Figure 6), suggesting a direct correlation between cholesterol levels and coronary atherosclerosis in this mouse model of CAD. Lipid content, plaque distribution, and the magnitude of stenosis was similar between the 2 groups of mice. TAC-operated Ldlr–/– mice that died during the physical stress protocol showed evidence of MI due to coronary occlusions (Supplemental Figure 7).
Although further characterization is necessary to define the causes of MI, the finding that afterload leads to the development of coronary lesions in Ldlr–/– mice is relevant, because this pathological model of CAD can be applied to complex genetically modified mice by using PCSK9 AAV technology (61).

In conclusion, the coexistence of high cholesterol and increased afterload can induce the formation of coronary lesions in approximately 95% of ApoE–/– mice and leads to plaque disruption and occlusion in more than 70% of the mice (Figure 7). This pathological phenotype recapitulates major complications of human coronary atherosclerosis, specifically the formation in mice of coronary lesions that can progress to disruption, leading to thrombosis or occlusion, ultimately causing myocardial events.

This study presents for the first time to our knowledge a reliable tool to investigate not only the pathogenesis of coronary atherosclerosis and its progression in ApoE–/– and Ldlr–/– mice, but also potential therapeutic targets.

**Methods**

**Minimally invasive TAC.** WT C57BL/6J, ApoE–/–, and Ldlr–/– male mice (11–14 weeks of age; body weight 25–27 g) were subjected to TAC or sham operation (62). Briefly, mice were anesthetized using a single i.p. injection of ketamine (100 mg/kg) and xylazine (5 mg/kg). A topical depilatory was applied to the chest, and the area was cleaned with betadine and alcohol. A horizontal incision 0.5–0.8 cm in length was made at the second intercostal space. After retracting the thymus, the aortic arch was visualized with a dissecting scope (Zeiss Discovery.V8 Stereo) at low magnification. A 7-0 nylon ligature was tied between the innominate and left common carotid arteries with an overlying 28-gauge needle, which was then rapidly removed, leaving a discrete region of stenosis. In sham-operated animals, the ligature was just tied loosely around the aorta.

Transstenotic gradient was measured in TAC-operated ApoE–/– mice subjected to a 28-gauge needle ligation by using Telemetry catheters (HD-X10; Data Sciences International). Briefly, 1 day after TAC, mice were anesthetized using a single i.p. injection of ketamine (100 mg/kg) and xylazine (5 mg/kg). A gel-filled catheter connected to a blood pressure transducer was inserted first into the right carotid (high pressure) and secured with a 7-0 silk suture. The transducer, which communicates with the receiver wirelessly, recorded blood pressure continuously using the Ponemah Physiology Platform (Data Sciences International). Values were recorded for 10 minutes at 1-minute intervals, each reading representing the average of blood pressure during 10-second sampling period. The catheter was then inserted into the left common carotid artery and secured with a 7-0 silk suture. Blood pressure was recorded for 10 minutes, and the animals sacrificed afterward. The transaortic gradient was expressed as the difference in blood pressure (systolic and mean) upstream and downstream of the banding.

**Cholesterol levels.** Total cholesterol was measured in plasma from sham- or TAC-operated WT and ApoE–/– mice, using a Cholesterol E kit (W AKO Life Science) according to the manufacturer’s instructions. Briefly, whole blood was collected in EDTA via cardiac puncture. Plasma were obtained by centrifugation at 1,000 g for 15 minutes at 4°C.

**Stress-induced MI protocol.** Starting from 8 weeks after TAC, ApoE–/– mice were placed on the treadmill (Columbus Instrument) to run for 4 min/d, once a week, for 12 weeks, at a maximal sustained running speed of 13–14 m/min with 15° of inclination. Because this test is a form of forced exercise, when the mice sometimes stopped for short periods, they were gently tapped or lifted to encourage resumption of running. A minimal electric shock was also applied in order to encourage the run on the treadmill.

**Radiotelemetry for electrocardiographic analysis.** ECGs in conscious mice were measured using telemetry transmitters (ETA-F10 transmitter; Data Sciences International). Briefly, the transmitter device, which communicates with the receiver wirelessly, was inserted subcutaneously through a dorsal incision in the abdomen of the animal, and the 2 ECG electrodes were placed subcutaneously along the superior right pectoral region (negative lead) and the left caudal rib position (positive lead) to approximate lead II of the Einthoven surface ECG. Mice were housed individually in a temperature- and humidity-controlled facility and maintained under a 12-hour dark and 12-hour light cycle. They were allowed to recover for 7 days after surgery. ECG traces were continuously recorded at baseline, during the stress-induced MI protocol and during the following 48 hours, using LabChart software (AD Instruments).

**Echocardiographic analysis.** Cardiac dimensions and function were analyzed by transthoracic echocardiography using a Vevo 770 Imaging System (VisualSonics) as previously described (62). Mice were lightly anesthetized with inhaled isoflurane (0.2% in O2). Left ventricle M-mode was used, all measurements
were obtained from 3–6 consecutive cardiac cycles, and the average values were used for analysis. Left ventricle end-diastolic (LVDd) and end-systolic (LVDs) dimensions were measured from the M-mode traces, and fractional shortening was calculated as follows: \((\text{LVDd} - \text{LVDs}) / \text{LVDd}\). Diastolic measurements were taken at the point of maximum cavity dimension and systolic measurements were made at the point of minimum cavity dimension, using the leading-edge method of the American Society of Echocardiography (63).

**Oil Red O staining.** To assess lipid deposition in the coronary arteries, frozen heart sections were stained with Oil Red O (MilliporeSigma) as previously described (64). Briefly, heart cryosections were washed in PBS, stained with 0.2% Oil Red O in 60% isopropanol for 20 minutes, and washed with 60% isopropanol. After counterstaining performed with Mayer's hematoxylin, sections were washed and coverslipped with aqueous mounting medium. Images were acquired by using a Zeiss Axio Observer.Z1 microscope, and the area of Oil Red O staining was calculated by using ImagePro (Media Cybernetics). Coronary stenosis was calculated by measuring the area of the lumen \((A_L)\) and the internal elastic lamina \((A_{IEL})\); the percentage of stenosis was calculated as follows: \((A_{IEL} - A_L) / A_{IEL} \times 100\). Oil Red O percentage in the plaque stenosis was calculated as the ratio of positive Oil Red O staining area to the percentage plaque area.

**Masson's trichrome staining.** Myocardial fibrosis was assessed by TAC-operated WT and ApoE−/− mice by using Masson's trichrome stain kit (Polysciences Inc.) according to the manufacturer's instructions.

**Histology and immunostaining.** Mouse hearts were collected no more than 1 hour postmortem without perfusion and fixed with 4% paraformaldehyde (PFA) overnight at 4°C. The following day, 4% PFA was replaced by 30% sucrose and left overnight at 4°C on rotation. The hearts were divided into 3 parts (base, center, and apex), submersed overnight with OCT/sucrose 30% (1:1 ratio), and consequently embedded in OCT. The whole heart was sectioned, with sets of 10 consecutive sections of 10 μm collected every 30 μm. Heart sections every 260 μm were routinely stained with H&E and coverslipped with aqueous mounting medium. For immunofluorescence analysis, the heart sections were permeabilized in 0.5% Triton X-100 in a solution of 5% BSA 5%, followed by blocking in BSA 5% (45 minutes at room temperature). Next, the heart sections were incubated overnight at 4°C with anti-CD41 antibody (1:100, 553897; BD Pharmingen), anti-α smooth muscle actin antibody (1:200, 180186; Life Technologies), anti-CD31 (1:100, 553370; BD Biosciences), anti-CD68 antibody (1:200, MCA1957; Bio-Rad), and anti–collagen I antibody (1:200, ab43710; Abcam) in 5% BSA. After 3 washes in PBS, secondary antibodies Alexa Fluor 568–conjugated donkey anti-mouse IgG (1:200, ab34710; Abcam), Alexa Fluor 647–conjugated goat anti-rat IgG (1:500, A21247, Thermo Fisher Scientific), and Cy5-labeled donkey anti-rabbit IgG (1:500, 711-175-152; Jackson ImmunoResearch) were added for 1 hour at room temperature. Nuclei were counterstained with Hoechst 33258 (1:10,000, 382061; Calbiochem). Immunofluorescence images of heart sections were captured in a Z-stack with 0.48-μm steps using a Zeiss Axio Observer.Z1 microscope followed by deconvolution using the ZEN pro 2012 software (Zeiss).

**Electron microscopy.** A 1-mm thin tissue was collected from the anterior wall of the left ventricle, showing a pale area. We used hearts of ApoE−/− TAC-operated mice in which ECG traces showed ST elevation and control hearts from C57BL/6 mice at the same time points after TAC. Cardiac left ventricle tissues were fixed with a modified Karnovsky’s fix (2.5% glutaraldehyde, 4% PFA, and 0.02% picric acid in 0.1 M sodium cacodylate buffer at pH 7.2) and post-fixed in 1% OsO4 and 1.5% K2[Fe(CN)]6. After dehydration, samples were embedded in an Epon analog resin. Ultrathin sections were cut using a Diatome diamond knife (Diatome) on a Leica Ultracut S ultramicrotome (Leica). Sections were collected on copper grids and stained with uranyl acetate and lead citrate and then visualized on a JEM 1400 electron microscope (JEOL) operated at 100 kV. Images were recorded with a Veleta 2 K × 2 K digital camera (Olympus-SIS).

**Statistics.** Statistical analysis was performed with GraphPad Prism 7.0 software. All data are reported as mean ± SEM. Statistical analysis was performed with 1-way ANOVA followed by Dunnett’s post-hoc test when comparing more than 2 groups of data (Figure 1H and Figure 2G) and by unpaired t test when comparing 2 groups of data (Supplemental Figure 1). Data were considered statistically significant when a value of at least \(P < 0.05\) was achieved.

**Study approval.** All of the studies were performed according to protocols approved by the Weill Cornell Institutional Animal Care and Use Committee.
Author contributions
ADL, AM, and YZ designed the studies. AM, YZ, LR, MAR, and JEI conducted experiments and acquired data. ADL, AM, YZ, MAR, and JEI analyzed data. ADL and AM wrote the manuscript. JEI contributed to the editing of the manuscript.

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